

**TITLE OF THE INVENTION: Method for the screening of  $\alpha_2\delta$ -1 subunit binding ligands**



5 This application is a Continuation of USSN 09/397,549 filed September 16, 1999; the entire contents of which are herein incorporated by reference.

**FIELD OF THE INVENTION**

The invention relates to a method for the screening of ligands which bind a soluble secreted cerebral cortical voltage-dependent calcium channel  $\alpha_2\delta$ -1 subunit polypeptide.

10 **BACKGROUND OF THE INVENTION**

Gabapentin (1-aminoethyl-cyclohexane acetic acid) is currently commercialized for the treatment of epilepsy. The compound has however been recognized as being also useful for the treatment of pain and anxiety.

15 Recent reports have suggested an interaction between gabapentin and the  $\alpha_2\delta$  subunit of a voltage-dependent calcium channel (VDCC). But electro-physiological studies have yielded conflicting data on the action of gabapentin at VDCCs, even though the relevance of the interaction of gabapentin at the  $\alpha_2\delta$  subunit to the clinical utility of the drug is becoming clearer. However, none of the prototype anticonvulsant drugs displace  
20 [ $^3\text{H}$ ]gabapentin binding from the  $\alpha_2\delta$ -1 subunit.

The most frequently used assay currently available for the screening of ligands that bind the  $\alpha_2\delta$  subunit involves the use of pig membrane extracts as a source of the  $\alpha_2\delta$  subunit. Such an assay presents major inconvenience. Firstly, because the assay material is a  
25 membrane extract, it is very difficult to accurately determine the protein composition from one assay preparation to another particularly with regard to the subtype. Also, the presence of various impurities in the assay preparation is a problem in small plate assays. Furthermore, as the protein preparation lacks homogeneity, the interaction between the targeted protein and the assay plate is often quite uneven. This renders the streamlining of  
30 the assay in a high throughput format almost impossible to achieve.

Utility Application

### SUMMARY OF THE INVENTION

The inventors have found that it was possible to use a soluble secreted form of a voltage-  
5 dependant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide (hereinafter  $\alpha_2\delta$ -1 subunit polypeptide) in an assay for the screening of ligands which bind the  $\alpha_2\delta$ -1 subunit.

The exact position and configuration of the [ $^3\text{H}$ ]gabapentin binding site on the  $\alpha_2\delta$  subunit is not currently known. Furthermore, recent deletion experiments on the porcine  $\alpha_2\delta$ -1  
10 subunit coding sequence have shown that amino-acids close to the C-terminal region are needed in order for the protein to bind [ $^3\text{H}$ ]gabapentin. For this very reason, the use of truncated forms of the porcine  $\alpha_2\delta$ -1 subunit in screening assays has not been disclosed or suggested in the prior art because there was concern as to whether relevant levels of binding capacity would be achieved in an assay environment.

15 The assay of the invention is of considerable interest because it confirms that a recombinant soluble secreted  $\alpha_2\delta$ -1 subunit polypeptide can be used in high throughput  $\alpha_2\delta$ -1 ligand screening. It also provides a useful advantage over the pig membrane extract screening assay as it allows the study of  $\alpha_2\delta$ -1 subtype-specific binding ligands. Proteins can be tagged which makes purifying convenient and possible to use a tagged antibody for  
20 recognition.

It was not clear whether the addition of the 6His tag to the C-terminus of the protein would affect the [ $^3\text{H}$ ]gabapentin binding properties of  $\alpha_2\delta$

It was also unclear whether a C-terminally located 6His tag on  $\alpha_2\delta$  would be accessible for  
25 interaction with the Ni NTA chromatography matrix (for purification purposes) and SPA bead, or Ni flashplate well surface (for purposes of the assay).

The invention concerns a method for the screening of ligands which bind a calcium channel  $\alpha_2\delta$ -1 subunit.

30 The method comprises the steps of:

- contacting a secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide with:

- a ligand of interest; and
- a labelled compound which binds a  $\alpha_2\delta$ -1 subunit; and
- measuring the level of binding of the labelled compound to the secreted soluble  $\alpha_2\delta$ -1 subunit.

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The invention also concerns a kit for the screening of ligands which bind a calcium channel  $\alpha_2\delta$ -1 subunit.

The kit comprises:

- a secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide; and
- a labelled compound which binds a calcium channel  $\alpha_2\delta$ -1 subunit.

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#### **BRIEF DESCRIPTION OF THE FIGURES**

- 15 Figure 1 represents the elution profile of the recombinant polypeptide with the amino acid sequence of SEQ ID No 9 purified by Superdex-200 chromatography, either before or after electron on NI-NTA.

- Figure 2 illustrates the optimization of imidazole concentrations in an embodiment of the  
20 SPA assay of the invention.

Figure 3 illustrates the optimization of imidazole concentrations in an embodiment of the flashplate assay of the invention.

- 25 Figure 4 illustrates the flashplate time course of [ $^3\text{H}$ ]gabapentin binding to various concentrations of the recombinant polypeptide with the amino acid sequence of SEQ ID No 9.

- Figure 5 illustrates the capacity of the recombinant polypeptide with the amino acid  
30 sequence of SEQ ID No 9 in a flashplate assay after 3 hours of incubation.

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Figure 6 illustrates the optimum imidazole concentration, assayed after 3 hours of incubation, required to maximize [ $^3\text{H}$ ]gabapentin binding using a constant amount of the recombinant polypeptide with the amino acid sequence of SEQ ID No 9.

- 5 Figure 7 illustrates flashplate assay of [ $^3\text{H}$ ]gabapentin saturation binding to the purified recombinant polypeptide with the amino acid sequence of SEQ ID No 9, assayed after 3 hours of incubation.

- Figure 8 illustrates the flashplate time course optimisation of imidazole concentration required to maximize the [ $^3\text{H}$ ]Leucine binding window to to the purified recombinant  
10 polypeptide with the amino acid sequence of SEQ ID No 9, assayed after 3 hours of incubation.

Figure 9 illustrates competition curves of three compounds in the flashplate assay format, assayed after 3 hours of incubation.

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#### **DETAILED DESCRIPTION OF THE INVENTION**

- The invention concerns a method for the screening of ligands which bind a soluble secreted  $\alpha_2\delta$ -1 subunit polypeptide. The term  $\alpha_2\delta$ -1 subunit polypeptide, when used herein, is  
20 intended to designate a structure containing two polypeptides ( $\alpha_2$  and  $\delta$ ) attached to one another by covalent disulfide bridges. More particularly, the targeted  $\alpha_2\delta$ -1 subunit binding site is preferably the [ $^3\text{H}$ ]gabapentin binding site. The various parameters of the method of the invention are described in further detail below.

#### **25 A – Secreted soluble recombinant $\alpha_2\delta$ -1 subunit polypeptide**

- Several nucleotide sequences encoding a secreted soluble form of an  $\alpha_2\delta$ -1 subunit can be used in the context of the present invention. Preferred soluble secreted  $\alpha_2\delta$ -1 subunit polypeptides are derived from eukaryotic  $\alpha_2\delta$ -1 subunits, more preferably from mammal, such as mouse, rat, rabbit, porcine, bovine or others and human  $\alpha_2\delta$ -1 subunits. Most  
30 preferred soluble secreted  $\alpha_2\delta$ -1 subunit polypeptides are derived from the human or porcine  $\alpha_2\delta$ -1 subunits.



More specifically, the selected nucleotide sequences encode a secreted soluble polypeptide having at least 80%, preferably 90%, more preferably 95%, and most preferably 98 or 99% amino-acid identity with the polypeptide comprising from amino acid 1 to between amino-acids 985 and 1054, preferably between amino-acids 985 and 1059, and most preferably between amino-acids 1019 and 1044 of SEQ ID NO:5 or SEQ ID NO:16.

In order to determine the optimal deletions on the  $\alpha_2\delta$ -1 subunit cDNA that yield a soluble secreted polypeptide devoid of membrane anchorage structures and having a functional [ $^3\text{H}$ ]gabapentin binding site, the inventors tested the expression of several human or porcine  $\alpha_2\delta$ -1 subunit cDNA deletion mutants. The discussion provided below provides detailed comments on possible truncations, giving as an example the porcine  $\alpha_2\delta$ -1 subunit. However, given the very substantial cross-species homology for  $\alpha_2\delta$ -1 subunit sequences, the comments below can also be applied to other eukaryotic species, and more particularly other mammalian species such as the rat, the mouse or the rabbit. Their  $\alpha_2\delta$ -1 subunit sequences, which are available in public databases, share a very substantial homology with the human and porcine  $\alpha_2\delta$ -1 subunit sequences.

The inventors found that by deleting from the porcine  $\alpha_2\delta$ -1 subunit cDNA a nucleotide sequence encoding as much as amino-acids 967 to 1091 of the native protein, soluble polypeptides could be obtained. On the other hand, the minimal deletion required to achieve solubility appears to be located around nucleotides encoding amino-acids 1064 to 1091 of the sequence of SEQ ID NO:5. In this regard, the mutant polypeptide expressed using a cDNA deletion mutant from which a sequence encoding amino-acids 1064 to 1091 is removed is found in both soluble and membrane-associated forms, with [ $^3\text{H}$ ]gabapentin and/or other derivatives or compounds such as pregabalin and gabapentoids binding properties similar to that of the wild type protein. Furthermore, a mutant protein expressed using a cDNA deletion mutant from which a nucleotide sequence encoding amino-acids 1085 to 1091 is removed recovers its membrane anchorage properties. Also, mutant proteins expressed using cDNA deletion mutants from which nucleotide sequences encoding either amino-acids 1037 to 1091 or amino-acids 1019 to 1091 of SEQ ID NO:5 or 16 are removed are found in soluble form.

The inventors believe that the soluble secreted  $\alpha_2\delta$ -1 subunit polypeptides which are as close as possible to the native sequence and which are therefore more likely to retain their native folding and hence their [ $^3\text{H}$ ]gabapentin- binding properties are those corresponding to a protein in which amino-acid stretch 985-1091 to 1074-1091, the amino-acid sequence of SEQ ID NO:5 or 16 has been deleted. The skilled scientist can quite easily determine within this amino-acid stretch the optimal mutant protein.

The invention therefore particularly concerns a screening assay in which the secreted soluble  $\alpha_2\delta$ -1 subunit polypeptide is preferably a polypeptide having at least 80% identity with the polypeptide comprising from amino-acid 1 to between amino-acid 985 and 1054, preferably between amino-acids 985 and 1059, and most preferably between amino-acids 1019 and 1064 of SEQ ID NO:5 or SEQ ID NO:16. Preferred  $\alpha_2\delta$ -1 subunit polypeptides which can be used in the present invention are those of SEQ ID N°6, 7, 8, 9, 13, 14 and 15, with the polypeptides of SEQ ID NO:9 or SEQ ID NO:15 being most preferred.

In a first and preferred embodiment of the invention, the  $\alpha_2\delta$ -1 subunit polypeptide is purified before it is used in the assay. The purification step, an example of which is provided further in this specification, can be carried out using several purification techniques well-known to the skilled person.

In some instances, it is required to tag the  $\alpha_2\delta$ -1 subunit polypeptide prior to purification. The tag is then in most instances encoded into the nucleotide sequence that is needed to express the polypeptide. Examples of such tags include, but are not limited to sequences encoding C-myc, FLAG, a sequence of histidine residues, heamagglutinin A, V5, Xpress or GST. Most of these tags can be incorporated directly into the sequence, for instance through PCR amplification by incorporating the appropriate coding sequence in one of the PCR amplification primers. However, the tag can also be introduced by other means such as covalent binding of the appropriate nucleic acid sequence encoding the tag moiety with the 5' or 3' end of the nucleic acid sequence encoding the polypeptide sequence. This is the

case for GST. It should be noted that the tag can be located at either end of the polypeptide sequence. Furthermore, in some instances, it can be advantageous to insert a cleavage site between the tag and the polypeptide sequence in order to permit removal of the tag sequence if needed.

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In other cases, providing a tag to the polypeptide is not needed. For instance, the protein can be purified using affinity columns loaded with specific monoclonal antibodies.

10 In a second embodiment of the invention, the  $\alpha_2\delta$ -1 subunit polypeptide can be only partially purified. For instance, it can be purified along with other contaminating proteins using an appropriate chromatography matrix such as ion-exchange chromatography column. In such instances, it is not required to tag the desired polypeptide of interest.

15 The most preferred embodiment contemplated by the inventors concerns the use of a purified tagged  $\alpha_2\delta$ -1 subunit polypeptide. A particularly preferred tag is a nucleotide sequence encoding from 2 to 10, and preferably 6 histidine residues as provided in the polypeptide of SEQ ID No 9.

20 With regard to the  $\alpha_2\delta$ -1 subunit polypeptide used subsequently in the screening assay of the invention, several possibilities are also open to the skilled person.

In a first and preferred embodiment, the  $\alpha_2\delta$ -1 subunit polypeptide comprises a tag moiety which can be selected among the tags referred to above. Such tagged polypeptides are particularly useful in SPA or flashplate assays. A preferred tag is the nucleotide sequence  
25 encoding histidine residues referred to above.

In a second embodiment, the  $\alpha_2\delta$ -1 subunit polypeptide can be used without a tag. This is the case for instance in SPA or flashplate assays comprising beads or plates coated with wheat germ lectin. In such an embodiment, the tag is not needed as the carbohydrate  
30 moieties of the  $\alpha_2\delta$ -1 subunit polypeptide bind directly to the wheat germ lectin-coated beads or plates.

**B - Labelled compounds which bind the  $\alpha_2\delta$ -1 subunit polypeptide**

In cases where the  $\alpha_2\delta$ -1 binding site is the [ $^3\text{H}$ ]gabapentin binding site, the preferred labelled compound which can be used is of course gabapentin itself. However, gabapentin  
 5 is not the only labelled compound which can be used in this context. Indeed, it has been previously demonstrated that saturation binding analyses on porcine synaptic plasma cerebral cortex membranes performed in the presence of L-leucine indicate a competitive interaction of the amino acid with the [ $^3\text{H}$ ]gabapentin binding site, significantly reducing [ $^3\text{H}$ ]gabapentin binding affinity for the site. The inventors believe that this competitive  
 10 interaction is true across across all the amino-acids listed in table 1 below.

**Table 1**

**Binding affinities of selected amino acids ( $\text{IC}_{50} < 500\text{nM}$ ) for the [ $^3\text{H}$ ]gabapentin site in  
 15 porcine cortical membranes**

	<b><u>COMPOUND</u></b>	<b><u><math>\text{IC}_{50}</math> (NM) ARITHMETIC MEAN (N=3) <math>\pm</math> S.E.M.</u></b>
	Gabapentin	$42.1 \pm 5.5$
	L-Norleucine	$23.6 \pm 6.7$
20	L-Allo-Isoleucine	$32.8 \pm 6.0$
	L-Methionine	$49.6 \pm 10.0$
	L-Leucine	$61.3 \pm 20.9$
	L-Isoleucine	$68.8 \pm 1.9$
	L-Valine	$330 \pm 18$
25	L-Phenylalanine	$351 \pm 89$

It is therefore possible to use commercially available labelled forms of these high affinity ligands in replacement of gabapentin. The utility of [ $^3\text{H}$ ]L-leucine has been demonstrated in a filter binding assay and in a flashplate assay format. The inventors believe that labelled  
 30 amino acids but also other compounds, with affinities preferably below 500 nM in the binding assay can be used as replacements of gabapentin.

With regard to the label, several embodiments can be used in the context of the invention. Preferred labels are of course radioactive labels, a list of which is provided further in this specification.

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#### **C - Assay formats and conditions**

Several assay formats can be used to carry out the method of the present invention. Preferred assay formats include scintillation assays such as the scintillation proximity assay (SPA) or the flashplate assay. Other assay formats well known to those skilled in the arts such as the filter binding assay and the centrifugation assay are also contemplated in the present invention.

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SPA and flashplate assays are preferred assay formats for the present invention. Additional details on these assays are provided below.

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#### **Scintillation assay format**

Scintillation assays technology either involves the use of scintillant beads (for the SPA assay) or plates (for the flashplate assay). SPA beads are usually made from either cerium-doped yttrium ion silicate ( $\text{Y}_2\text{SiO}_5\text{:Ce}$ ) or polyvinyltoluene (PVT) containing an organic scintillant such as PPO. Flashplates commonly used are those such as Ni chelate flashplates although other flashplates can also be used.

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Assays are usually carried out in aqueous buffers using radioisotopes such as  $^3\text{H}$ ,  $^{125}\text{I}$ ,  $^{14}\text{C}$ ,  $^{35}\text{S}$  or  $^{33}\text{P}$  that emit low-energy radiation, the energy of which is easily dissipated in an aqueous environment. For example, the electrons emitted by  $^3\text{H}$  have an average energy of only 6 keV and have a very short path length ( $\sim 1\text{ }\mu\text{m}$ ) in water. If a molecule labelled with one of these isotopes is bound to the bead or flashplate surface, either directly or via interaction with another molecule previously coupled to the bead or flashplate, the emitted radiation will activate the scintillant and produce light. The amount of light produced, which is proportional to the amount of labelled molecules bound to the beads, can be measured conveniently with a liquid scintillation (LS) counter. If the labelled molecule is

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not attached to the bead or a flashplate surface, its radiation energy is absorbed by the surrounding aqueous solvent before it reaches the bead, and no light is produced. Thus, bound ligands give a scintillation signal, but free ligands do not, and the need for a time-consuming separation step, characteristic of conventional radioligand binding assays, is eliminated. The manipulations required in the assays are reduced to a few simple pipetting steps leading to better precision and reproducibility.

The conditions under which SPA and flashplate assays are performed in the context of the present invention are provided below.

## 10 Scintillation assay conditions

### 1) SPA assay

The SPA assays is first developed to optimize the conditions under which the radioligand binds the  $\alpha_2\delta$ -1 subunit polypeptide. The parameters which can be varied to optimize radioligand binding in a typical SPA assay using Amersham beads include assay temperature,  $\alpha_2\delta$ -1 subunit polypeptide interaction with the radioligand and the SPA beads, radioligand concentration as well as pH variations.

The temperature at which the assay can be carried out can vary from 1 to 30°C. Preferred temperatures range from 18 to 23°C, with 21°C being the most preferred temperature. The interaction of the  $\alpha_2\delta$ -1 subunit polypeptide with the SPA beads can be optimized by adjusting the concentration of the polypeptide and by introducing a reagent which will favor this interaction. When 50 mg of Amersham SPA beads are used, the  $\alpha_2\delta$ -1 subunit polypeptide concentration may vary from 0.1 to 10 pmoles per well, with the optimal concentration being generally around 5 to 6 pmoles per well.

As for the reagent favoring the interaction between the  $\alpha_2\delta$ -1 subunit polypeptide and the radioligand as well as the Amersham SPA beads, the inventors found that imidazole could be efficiently used for that purpose when the  $\alpha_2\delta$ -1 subunit polypeptide is tagged with an amino acid sequence including 6 histidine residues. Furthermore, and more importantly, it was found that imidazole also enhanced binding of the radioligand to the  $\alpha_2\delta$ -1 polypeptide.

The optimal concentration of imidazole used to enhance radioligand binding varies depending on the concentration of  $\alpha_2\delta$ -1 subunit polypeptide used in the assay. For instance, when the concentration of the  $\alpha_2\delta$ -1 subunit polypeptide is about 20  $\mu$ l ( $\alpha_2\delta$ -1 polypeptide concentration of 0.6 pmol/ $\mu$ l), imidazole concentrations ranging from 10 to 50 mM can be used, with concentrations ranging between 10 and 30 mM being preferred. A most preferred imidazole concentration is 20 mM. It is to be noted that other compounds such as histidine can be used to enhance radioligand binding. Furthermore, pH variations can also influence radioligand binding although pH variations should be closely monitored as they may have an effect on the structural configuration of the of  $\alpha_2\delta$ -1 subunit polypeptide. Also the use of imidazole is preferred to enhance radioligand binding, the person skilled in the art know that the use of imidazole is preferred but is absolutely not essential.

The concentration of the radioligand is evaluated with respect to the concentration of  $\alpha_2\delta$ -1 subunit polypeptide present in the assay medium. Generally, the concentration of radioligand varies from 1 nM to 100 nM. A preferred [ $^3$ H]gabapentin concentration is about 5 to 20 nM, with a most preferred concentration being about 10 nM. A preferred [ $^3$ H]leucine concentration is also about 5 to 20 nM, with a most preferred concentration being about 10 nM. It is to be noted that the concentration of other radioligands having affinities similar to those of [ $^3$ H]gabapentin and [ $^3$ H]leucine should also be in the range of about 5 to 20 nM.

Once the optimal radioligand binding conditions have been determined, a test ligand can be introduced in the assay medium to evaluate the level of displacement of the radioligand. The concentration of test ligand to be introduced in the assay medium usually varies from 0.1 nM to about 100  $\mu$ M. A preferred test ligand concentration of about 10  $\mu$ M is usually a starting point in a high throughput screening assay. Then, depending on the number of hits obtained, it may be lowered or increased.

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It is to be noted that the parameters set forth above, which have been evaluated for a typical SPA assay using Amersham SPA beads can be adjusted by the skilled person, for example if SPA beads of a different type are used.

5    **2) Flashplate assay**

Similarly to the SPA assays, the flashplate can first be developed in order to optimize the conditions under which the radioligand binds the  $\alpha_2\delta$ -1 subunit polypeptide. The parameters which can be varied to optimize radioligand binding in a typical flashplate assay using NEN Ni chelate flashplates also include assay temperature,  $\alpha_2\delta$ -1 subunit  
10    polypeptide interaction with both the radioligand and the flashplates, radioligand concentration as well as pH variations.

The temperature at which the assay can be carried out can vary from 1 to 30°C. Preferred temperatures range from 18 to 23°C, with 21°C being the most preferred temperature.

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The interaction of the  $\alpha_2\delta$ -1 subunit polypeptide with the flashplates can be optimized by adjusting the concentration of the polypeptide and by introducing a reagent which will favor this interaction. When a standard NEN Ni chelate flashplate is used, the  $\alpha_2\delta$ -1 subunit polypeptide volume usually varies between 0.5 and 20  $\mu$ l for a concentration of  
20     $\alpha_2\delta$ -1 subunit polypeptide of 0.6 pmol/ $\mu$ l. As the published maximum binding capacity of NEN plates is about 6 pmol per well, the inventors consider that an optimal concentration of  $\alpha_2\delta$ -1 subunit polypeptide is probably around 5 pmol per well at 8 $\mu$ l.

Also the use of imidazole is preferred to enhance radioligand binding, the person skilled in  
25    the art know that the use of imidazole is preferred but is absolutely not essential.

With regard to the reagent favoring the interaction between the  $\alpha_2\delta$ -1 subunit polypeptide and the radioligand as well as the flashplates, the inventors found that imidazole could also be efficiently used for that purpose when the  $\alpha_2\delta$ -1 subunit polypeptide is tagged with an amino acid sequence including 6 histidine residues. It was also found that imidazole  
30    concentrations substantially enhanced binding of the radioligand to the  $\alpha_2\delta$ -1 polypeptide. The optimal concentration of imidazole used to enhance radioligand binding varies



depending on the concentration of  $\alpha_2\delta$ -1 subunit polypeptide used in the assay. For instance, when the volume of the  $\alpha_2\delta$ -1 subunit polypeptide is about 10  $\mu$ l (  $\alpha_2\delta$ -1 polypeptide concentration of 0.6 pmol/ $\mu$ l), the optimal imidazole concentration can vary between 1 and 20 mM, with a concentration of about 10 mM being preferred. As  
5 mentioned previously, other compounds such as histidine as well as pH variations may be used to enhance radioligand binding.

The concentration of the radioligand is evaluated with respect to the concentration of  $\alpha_2\delta$ -1 subunit polypeptide present in the assay medium. Generally, the concentration of  
10 radioligand varies from 1 nM to 100 nM. A preferred [ $^3$ H]gabapentin concentration is about 5 to 20 nM, with a most preferred concentration being about 10 nM. A preferred [ $^3$ H]leucine concentration is also about 5 to 20 nM, with a most preferred concentration being about 10 nM. It is to be noted that the concentration of other radioligands having affinities similar to those of [ $^3$ H]gabapentin and [ $^3$ H]leucine should also be in the range of  
15 about 5 to 20 nM.

Once the optimal radioligand binding conditions have been determined, a test ligand can be introduced in the assay medium to evaluate the level of displacement of the radioligand. The concentration of test ligand to be introduced in the assay medium usually varies from  
20 0.1 nM to about 100  $\mu$ M. A preferred test ligand concentration of about 10  $\mu$ M is usually a starting point in a high throughput screening assay. Then, depending on the number of hits obtained, it may be lowered or increased.

The inventors have tested the displacement of a particular radioligand, [ $^3$ H]gabapentin, with (S+)-3-isobutyl gaba, (R-)-3-isobutyl gaba and gabapentin. The data provided in the  
25 examples which follow clearly shows that the assay can be used in high throughput competition studies.

**Example 1****Construction of a nucleotide sequence encoding the putative soluble porcine  $\alpha_2\delta$ -1b deletion mutant of SEQ ID NO:9****a) Primer design**

- 5 PCR primers were designed to generate the soluble porcine  $\alpha_2\delta$ -1b deletion mutant of SEQ ID NO:9 as follows:

5' PCR primer: This was designed to engineer in a KOZAK translation initiation consensus sequence prior to the coding sequence (Kozak *JBC* 266 19867-19870)

- 3' PCR primer: This was designed to engineer in six histidine residues followed by a stop-codon at the desired location in the coding sequence. In addition to the stop codon the  $\alpha_2\delta$ -1 primers also included an *Eco* RI restriction site.

- The bold region in each primer sequence denotes the 'tagged' region; addition of sequences not present in the template. Primers were custom synthesized by Perkin Elmer Applied Biosystems UK to the ABI ready pure grade, supplied lyophilized then resuspended to 15  $\mu$ M in 10mM TE. JB189 and 195 were provided without 5' phosphate groups:

5' primer JB189 (5'-TCGCCACCATGGCTGCTGGCTGCCTGCTG-3', SEQ ID NO:20)

- 20 3' primer JB195 (5'-TCGGAATTCCTCAGTGATGGTGATGGTGATGAGAAACACCACCACAGTCGGT-3', SEQ ID NO:21)

**b) PCR protocols for the generation of the  $\alpha_2\delta$ -1 deletion mutant**

- 25 **1) Generation of the pcDNA3-porcine- $\alpha_2\delta$ -(+) PCR template**

An oligo dT-primed  $\lambda$ gt10 porcine cerebral cortical cDNA library was screened by ECL (Amersham) using a 2,381-bp *HindIII* fragment (coding sequence 268-2649) of the rabbit skeletal muscle  $\alpha_2\delta$  clone (pcDNA3-Rab- $\alpha_2\delta$ -(+)) (supplied by Neurex) as the probe.

- A positive insert was identified and subcloned into pBluescript-SK-(+) to generate pB-PC- $\alpha_2\delta$ -1.1. The clone was sequenced on both strands, except for a 711-bp stretch at one end of the clone, which had a high degree of homology to mitochondrial C oxidase.

The  $\alpha_2\delta$  coding region was homologous to the 3' region of the human neuronal  $\alpha_2\delta$  sequence but lacked 926 bp of 5' coding sequence. The missing sequence was obtained by 5'-RACE using total RNA prepared from porcine cerebral cortex. RACE was performed across a *Bgl* I site unique in known  $\alpha_2\delta$  sequences (rabbit (accession no. M21948)), rat

5 (accession number M86621), human (accession no. M76559)

The sequence derived from the 5' RACE product was used to design a primer (JB042, 5'-GGGGATTGATCTTCGATCGCG-3'; SEQ ID NO:18) specific for the 5'-untranslated end of the cDNA. PCR was then performed with *Pfu* DNA polymerase using JB042 and a primer downstream of the *Bgl* I site (JB040, CTGAGATTGTTGGGGTTCTTTGG, SEQ ID

10 NO:19).

The PCR product was ligated to Eco RI linkers (5'-GGAATTCC-3') and then digested with Eco RI and *Bgl* I. The 1,564-bp fragment (5' portion of the  $\alpha_2\delta$  cDNA) was gel-purified.

15 Similarly, a 2,303-bp fragment (3' portion of the  $\alpha_2\delta$  cDNA) was isolated after digestion of pB-PC- $\alpha_2\delta$ -1.1 with *Bgl* I and Eco RI. The two fragments of  $\alpha_2\delta$  cDNA were then ligated to EcoRI-digested pcDNA3 in a three-way ligation. A clone was picked with the full-length  $\alpha_2\delta$  sequence in the positive orientation with respect to the cytomegalovirus promoter (pcDNA3-PC- $\alpha_2\delta$ -(+)).

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## 2) PCR protocol

The following reagents were added to obtain two cocktails labelled 'lower' and 'upper' buffers.

	<i>Lower</i>	$\mu$ l
25	10x <i>Pfu</i> DNA polymerase buffer	25
	10mM dNTP's	5
	100ng/ $\mu$ l pcDNA3-porcine- $\alpha_2\delta$ -(+)	10
	15 $\mu$ M JB189	8.5
	15 $\mu$ M JB195	8.5
30	H <sub>2</sub> O	193

<i>Upper</i>	$\mu$ l
10x <i>Pfu</i> DNA polymerase buffer	25
H <sub>2</sub> O	220
2.5units/ $\mu$ l <i>Pfu</i> DNA polymerase	5

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50 $\mu$ l aliquots of lower buffer were added to each of four 0.5ml eppendorf tubes. To each was added one PCRgem 100 ampliwx bead (PE biosystems). Tubes were heated to 80°C for 2 minutes then cooled to 4°C. 50 $\mu$ l of upper buffer was then added to each tube. Tubes were then cycled on a Stratagene Robo-Cycler according to the following conditions: 98°C / 1min 30sec, followed by: for 20 cycles 98°C / 45sec, 54°C / 2min, 72°C / 6min, followed by: 72°C / 20min, followed by: hold at 4°C.

The 3228bp PCR product was then purified on a QIAquick PCR purification column (Qiagen) and eluted with 61 $\mu$ l of H<sub>2</sub>O. The following reagents were added to the eluted

15 DNA: 0.7 $\mu$ l 10mM ATP, 7 $\mu$ l 10x Polynucleotide Kinase buffer, 1 $\mu$ l 1unit/ $\mu$ l Polynucleotide Kinase.

The above 5' phosphorylation reaction was incubated at 37°C for 1 hour. The reaction was stopped by incubation at 65°C for 10min. The 3228bp 5' phosphorylated PCR product was

20 then gel purified from a 1% agarose gel using QIAEX (Qiagen) beads and eluted in ~50 $\mu$ l.

## **Example 2**

### **Cloning of the PCR fragments of Example 1 into the Baculovirus transfer vector pFastBac1**

25 The PCR products of Example 1 (3228bp JB189/JB195 derived PCR product coding for 6His tagged porcine  $\alpha_2\delta$ -1b: SEQ ID No 9) were cloned into *Stu* I digested, calf intestinal phosphatase dephosphorylated, phenol chloroform extracted and QIAEX gel purified pFastBac1 (Life Technologies) using the Rapid DNA ligation kit (Roche Diagnostics) transforming XL1-blue ( $\alpha_2\delta$ -1b) *E. Coli* cells:

30

**a) Screening for positive recombinants**

- Given that the PCR product was cloned by blunt-end ligation a screen was required to select a recombinant with the gene ligated in the positive orientation with respect to the polyhedrin promoter in pFastBac1. This was achieved by restriction digest of miniprep
- 5 DNA (Qiagen miniprep kit) prepared from colony minicultures and analysis on a 1% TAE agarose gel. A positive clone was identified according to the following digest patterns:

SEQ ID No 9 in pFastBac1

*Eco* RI digest performed on miniprep DNA

10		Predicted fragments (bp)
	PCR product cloned in a positive orientation	4773 and 3230
	PCR product cloned in a negative orientation	7989 and 14

**b) Sequencing analysis of selected clones**

- 15 One positive was selected for this clone and used to prepare a plasmid DNA stock of the desired construct (QIAGEN maxi kit). Confirmatory sequence reactions were performed using the Big Dye terminator sequencing kit and run on an ABI 310 Prism Genetic Analyzer. Sequence analysis of both coding strands was performed using a selection of sequencing oligonucleotide primers and has yielded the following results:
- 20 Sequencing of pFBac-Porcine-s- $\alpha_2\delta$ -1- $\Delta$ 1040-1067-6His confirmed that the insert sequence corresponded to the nucleic acid encoding the polypeptide of SEQ ID No 9, except for the deletion of two bases from the 5' end of the 5' PCR primer (JB189). The loss of these two bases did not have any impact on the 5' end of the gene as the KOZAK
- 25 translation start-site consensus sequence (GCCACC) starts immediately after this deletion.

**Example 3****Protocol for establishing baculovirus banks for the expression of the  $\alpha_2\delta$ -1 deletion mutant of SEQ ID NO:9**

- 30 Essentially, the protocol used to generate the baculovirus banks is that outlined in the Life Technologies Bac-to Bac<sup>TM</sup> baculovirus expression systems manual.

**a) Transposition of DH10Bac *E. coli* cells**

- One ng (5µl) of the recombinant pFastBac-1 construct containing the nucleotide sequence encoding the porcine  $\alpha_2\delta$ -1 deletion mutant of SEQ ID No 9 was added to 100µl of
- 5 DH10Bac cells thawed on ice. The cells were then mixed gently by tapping the tube then incubated on ice for 30 minutes before heat shock treatment by incubation in a 42°C water bath for 45 seconds. The mixture was then chilled on ice for 2 minutes before the addition of 900µl of S.O.C. medium. The mixture was then placed in a shaking incubator (200rpm) at 37°C for 4 hours. The cells were then serially diluted (10 fold dilutions from  $10^{-1}$  to  $10^{-3}$ )
- 10 and 10µl of each dilution plated on LB agar plates containing 50µg/ml kanamycin, 7µg/ml gentamicin, 10µg/ml tetracycline, 100µg/ml Blue-gal and 40µg/ml IPTG. The plates were incubated at 37°C for between 1 and 3 days until discrete colonies of blue and white colour were discernible.

**b) Isolation of recombinant DNA**

- White colonies (containing the recombinant bacmid) were picked and grown for 24 hours (to stationary phase) at 37°C with shaking (200rpm) in 2ml of LB containing 50µg/ml kanamycin, 7µg/ml gentamicin and 10µg/ml tetracycline. 1.5ml of culture was then transferred to a microfuge tube and centrifuged at 14,000xg for 1 minute. The supernatant
- 20 was removed and the cells resuspended gently in 0.3ml of 15mM Tris-HCl (pH8.0), 10mM EDTA, 100µg/ml RNase A. 0.3ml of 0.2N NaOH, 1% SDS was then added and the mixture mixed gently before incubation at 22°C for 5 minutes. Then 0.3ml of 3M potassium acetate (pH5.5) was added and the sample placed on ice for 10 minutes. After centrifugation at 14,000xg for 10 minutes the supernatant was transferred to a tube
- 25 containing 0.8ml of isopropanol, mixed then placed on ice for 10 minutes before centrifugation at 14,000xg for 10 minutes. The supernatant was then discarded and the pellet rinsed with 0.5ml of 70% ethanol before centrifugation at 14,000xg for 5 minutes. This 70% ethanol rinse was then repeated before removing all of the supernatant and air drying the pellet for 10 minutes at room temperature. The pellet was finally resuspended in
- 30 40µl of TE.

**c) Transfection of sf9 cells with the recombinant bacmid DNA**

- A 6-well tissue culture plate was seeded with  $0.9 \times 10^6$  sf9 cells (cells at log phase having grown from a culture passaged at  $0.3 \times 10^6$  cells/ml) per 35mm well in 2ml of Sf-900 II SFM media containing 50units/ml penicillin and 50 $\mu$ g/ml streptomycin. Cells were left to
- 5 attach at 27°C for 1 hour. Bacmid DNA prepared as described above (5 $\mu$ l) was added to 200 $\mu$ l of Sf-900 II SFM media containing 6 $\mu$ l of CELLFECTIN and mixed before incubation at room temperature for 45 minutes. The cells were washed once with 2ml of Sf-900 II SFM media without antibiotics then 0.8ml of Sf-900 II SFM media was added to each tube containing the lipid-DNA complex. The wash buffer was removed from the cells
- 10 and the 1ml of diluted lipid-DNA complex overlaid on the cells. The cells were incubated for 5 hours at 27°C after which time the transfection mixture was removed and 2ml of Sf-900 II SFM media containing 50units/ml penicillin and 50 $\mu$ g/ml streptomycin was added. The cells were then incubated for 72 hours.
- 15 After incubation for 72 hours the media was removed from the cells and centrifuged at 500xg for 5 minutes. The supernatant was then transferred to a fresh tube, this was labelled as the P0 bank and stored at 4°C in the dark. The P1 bank was prepared by passaging sf9 cells at approx  $5 \times 10^6$  cells/ml to  $2 \times 10^6$  cells/ml (100ml in a 250ml Erlenmeyer flask) and adding 0.5ml of the P0 bank harvested above. The cells were then incubated shaking
- 20 (200rpm) at 27°C for 4 days. Under sterile conditions the culture was centrifuged at 500xg for 10 minutes and the supernatant 0.2 $\mu$ M filtered (P1 bank). The P2 bank was prepared by adding 2ml of P1 bank per 400ml culture (in 1L Erlenmeyer flasks) passaged as above to  $2 \times 10^6$  cells/ml. The culture was incubated as before for 4 days and the supernatant harvested and filtered as described for the P1 bank. The supernatant was first pooled then
- 25 aliquoted (10ml) and stored at 4°C.

**Example 4****Protein expression**

- To sf9 cells passaged from  $\sim 5 \times 10^6$  cells/ml to  $2 \times 10^6$  cells/ml in Sf-900 II SFM media was
- 30 added 0.1ml virus per 100 ml of cells of the appropriate viral bank (400ml volumes in 1L Erlenmeyer flasks). The cells were then cultured for 4-5 days at 27°C with 110 rpm

shaking. Expression of the protein was confirmed by SDS-PAGE and Western blotting using an anti penta-His monoclonal antibody (Qiagen) and was detected in the culture supernatant and cell lysate.

## 5 **Example 5**

### **Purification of $\alpha_2\delta$ -1 deletion mutant of SEQ ID NO:9**

The  $\alpha_2\delta$ -1 deletion mutant of SEQ ID NO:9 was purified from the cell lysate following the purification strategy outlined below:

10 The culture was centrifuged at 6,000xg for 10 minutes and the supernatant removed. The weight of the cell pellet was determined before re-suspension in 20mM Tris pH8.0, 100mM KCl, 1% P40-Nonidet (100ml per 20g of wet cells). A protease inhibitor cocktail (Sigma Cat# P8849), 1ml/L, was added to the mixture. The solution was then stirred for 10 minutes before centrifugation for 1 hour at 30,000xg and 4°C. The supernatant was concentrated (30kDa cut off) to approx. ~300ml then centrifuged for 1 hour at 100,000xg.

15

Supernatant containing the soluble proteins was diluted 1:3 in 10mM Tris-HCl pH8.0 (equilibration buffer) and loaded onto a pre-equilibrated Q-Sepharose column (2.5cm i.d. x 30cm h.) at a flow rate of 900ml/h. After washing with equilibration buffer until a stable  $A_{280nm}$  baseline had been achieved, protein was eluted with 20mM Tris-HCl pH8.0, 0.5M KCl, 10mM Imidazole.

20

The eluate was then loaded onto a Ni-NTA (Qiagen) column (2.5cm i.d. x 6cm h.) pre-equilibrated in 20mM Tris pH8.0, 0.5M KCl, 10mM Imidazole at a flow rate of 2 ml/min. The column was washed successively with buffer A (20mM Tris pH8.0, 0.5M KCl, 20mM Imidazole), buffer B (100mM Tris-HCl pH8.0, 1M KCl), and buffer A again. Elution was performed with buffer C (20mM Tris-HCl pH8.0, 100mM KCl, 0.5M Imidazole). The Ni-NTA eluate (~50ml) was concentrated (30kDa cut-off) to ~2ml and applied at 1ml/min and in 0.2ml aliquots, to an FPLC Superdex-200 column equilibrated in 10mM HEPES, pH7.4, 150mM NaCl. Fractions containing the polypeptide of SEQ ID No 9 were pooled. As shown in Figure 1, the protein elution profile and associated [ $^3$ H]gabapentin binding activity is presented together with a silver-stained SDS-PAGE gel (post Ni NTA load of

25

30



Superdex-200) demonstrating the co-elution of the ~130kDa band ( $\alpha_2\delta$ ) with the [ $^3\text{H}$ ]gabapentin binding activity and  $A_{280\text{nm}}$  profile.

### Example 6

#### 5 SPA assay of [ $^3\text{H}$ ]gabapentin binding to soluble porcine $\alpha_2\delta$ -1b-6His

The assay was carried out at 21°C. Assay components were added in the following order (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C) to 96-well Optiplates:

- 25 $\mu\text{l}$  imidazole at various concentrations (diluted from a 1M stock pH8.0, see assay details)
  - 10 50 $\mu\text{l}$  10mM HEPES pH 7.4
  - 25 $\mu\text{l}$  (50mg) SPA beads (Amersham)
  - 100 $\mu\text{l}$  s- $\alpha_2\delta$ -1b-6His of SEQ ID No 9 (2 $\mu\text{l}$  protein diluted to 100 $\mu\text{l}$ )
- obtained from example 5
- 25 $\mu\text{l}$  radioligand ([ $^3\text{H}$ ]gabapentin )

- 15 Immediately after adding radioligand, the optiplates were loaded in the Packard Top Count scintillation counter to follow the binding time course. Imidazole was first used in the assay to optimize the specific interaction of the protein's 6His tag with the SPA bead. Imidazole itself (up to 100mM) in the filtration assay has no effect on [ $^3\text{H}$ ]gabapentin binding (n=1).

20

As shown in figure 2, specific binding of [ $^3\text{H}$ ]gabapentin to the s- $\alpha_2\delta$ -1b-6His was enhanced by imidazole. Of the concentrations, tested the optimal was 50mM. Equilibration was reached after ~2hours.

#### 25 Example 7

#### Ni Flashplate assay of [ $^3\text{H}$ ]gabapentin binding to soluble porcine $\alpha_2\delta$ -1b-6His (SEQ ID No 9)

Assays were carried out at 21°C in a final volume of 250 $\mu\text{l}$  in 96-well NEN Ni chelate flash plates. Assay components were added in the following order (all reagents were

- 30 diluted in 10mM HEPES (pH 7.4 at 21°C)):

25 $\mu\text{l}$  10mM HEPES pH7.4

- 25µl imidazole at various concentrations (diluted from a 1M stock  
pH8.0, see assay details)  
75µl 10mM HEPES pH 7.4  
100µl s-α<sub>2</sub>δ-1b-6His (2µl protein diluted to 100µl) obtained from  
example 5  
25µl radioligand ([<sup>3</sup>H]gabapentin)

Immediately after adding the radioligand, flash plates were loaded in the Packard Top  
Count scintillation counter to follow the binding time course. The '[<sup>3</sup>H] flash plate'  
programme (cpm) was used to monitor activity. Imidazole was first used in the assay to  
optimize the specific interaction of the protein's 6His tag with the Ni flashplate. Imidazole  
itself (up to 100mM) in the filtration assay has no effect on [<sup>3</sup>H]gabapentin binding (n=1).

As shown in figure 3, the specific binding of [<sup>3</sup>H]gabapentin to the s-α<sub>2</sub>δ-1b-6His was  
enhanced by imidazole. Initially, from the concentrations tested, the best concentration was  
found to be 10mM.

Specific binding was determined at different volumes of s-α<sub>2</sub>δ-1b-6His, in the presence of  
10mM imidazole, over a time period of 10h. Results are shown in figure 4 and equilibrium  
was reached at ~3h. Specific binding increased linearly with increasing amounts of protein,  
up to 8µl, after which the binding capacity of the Ni chelate in the assay well was probably  
exceeded (see figure 5). The published maximum binding capacity of NEN plates is  
6pmol/well. The concentration of purified s-α<sub>2</sub>δ-1b-6His is estimated at ~0.6pmol/µl,  
which yields 5pmol/well at 8µl.

25

## **Table 2**

### **Saturation studies**

Saturation experiments were performed with 12 duplicate data points, [<sup>3</sup>H]gabapentin  
concentration ranged from ~1 to 350nM. Data was analyzed using KEL-RADLIG for  
Windows.

<u>Flash plate</u> (2µl protein used, n=2)	<u>Filter binding</u> K <sub>D</sub> (nm) (4µl protein used, n=3)
K <sub>D</sub> , 9.32nM K <sub>D</sub> , 10.5nM  <b>Mean = 9.91nM</b>	K <sub>D</sub> , 12.3nM K <sub>D</sub> , 8.91nM K <sub>D</sub> , 10.6nM  <b>Mean = 10.60 ± 0.98nM</b>

**Example 8****Ni Flashplate assay of [<sup>3</sup>H]Leucine binding to soluble porcine  $\alpha_2\delta$ -1b-6His**

The procedure described in example 2 was repeated, except that [<sup>3</sup>H]gabapentin was replaced by 25  $\mu$ l (10.1 nM) of [<sup>3</sup>H]Leucine, as shown in figure 8, [<sup>3</sup>H]Leucine binds with high affinity to soluble  $\alpha_2\delta$ -1b-6His. This demonstrates that it is possible to use commercially available forms of [<sup>3</sup>H]Leucine in replacement of [<sup>3</sup>H]gabapentin in the assay.

**Example 9**

10 **Ni Flashplate assay studying competitive binding of [<sup>3</sup>H]gabapentin, (S+)-3-isobutyl GABA and (R-)-3-isobutyl GABA to porcine  $\alpha_2\delta$ -1b-6His (SEQ ID No 9)**

Assays were carried out at 21°C in a final volume of 250 $\mu$ l in 96-well NEN Ni chelate flash plates. Wells were set up for both 'total' and 'non-specific' binding. Specific binding was defined as that remaining after subtraction of the average of the 'non-specific binding' values from the average of the 'total' binding values. Assay components were added in the following order (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C)):

	25 $\mu$ l	10mM HEPES pH7.4 or 25 $\mu$ l of the test compound at the appropriate concentration in HEPES
20	25 $\mu$ l	200 mM imidazole (diluted from a 1M stock pH8.0, see assay details)
Total binding	75 $\mu$ l	10mM HEPES pH 7.4
Non-specific binding	50 $\mu$ l	10mM HEPES pH 7.4 and 25 $\mu$ l 100 $\mu$ M (S+)-3-isobutyl GABA
25	100 $\mu$ l	s- $\alpha_2\delta$ -1b-6His (2 $\mu$ l protein* diluted to 100 $\mu$ l)
	25 $\mu$ l	radioligand ([ <sup>3</sup> H]gabapentin or [ <sup>3</sup> H]Leucine)

\* The source of s- $\alpha_2\delta$ -1b-6His was that purified by FPLC Superdex-200 gel filtration (see example 5)

30 Immediately after adding radioligand, flash plates were loaded in the Packard Top Count scintillation counter to follow the binding time course. Incubation time before the assay

was 3 hours. The '[<sup>3</sup>H] flash plate' programme (cpm) was used to monitor activity. Specific binding was ~98% of the 'total' value. Imidazole was used in the assay to optimize the specific interaction of the protein's 6His tag with the Ni flashplate. Imidazole itself (up to 100mM) in the filtration assay has no effect on [<sup>3</sup>H]gabapentin binding (n=1).

5

Competition studies were compared across the flash-plate and filter binding methodologies in order to validate the new assay technology with the established filter binding methodology.

- 10 GraphPad Prism software was used to process competition curve data and determine IC<sub>50</sub> and hill slope values. Twelve point competition curves with half log dilution steps of test compounds were used in the experiments. Results are shown in Table 3 below where IC<sub>50</sub> values are presented, and in figure 9 where hill slopes range from -0.9 to 1.3. The [<sup>3</sup>H]Gabapentin concentration used in assay is in the range of 10-13nM

15

### Table 3

#### Competition studies:

GraphPad Prism software was used to process competition curve data and determine IC<sub>50</sub> and hill slope values. Ten point competition curves with half log dilution steps of test

- 20 compounds were used in the experiments.

IC<sub>50</sub> values were converted to Ki values (presented in table) according to the following equation:

$$K_i = IC_{50} / (1 + [L]/K_D)$$

The K<sub>D</sub> values used were those mean values presented in table 1.

- 25 The [<sup>3</sup>H]Gabapentin concentration in the assay ranged from 10-13nM and was determined for each experiment for the purpose of calculating the Ki value as described above.

Hill slopes were all in the range of -0.9 to 1.3

<u>Test compound</u>	<u>Flash plate</u> (3µl protein used, n=2)	<u>Filter binding</u> $K_D$ (nm) (4µl protein used, n=3)
Gabapentin	10.4 7.97	7.13 7.70 10.2
Mean (geometric)	<i>9.10nM</i>	<i>7.84nM</i>
(S+)-3-isobutyl GABA	10.9 7.58	6.52 6.21 8.29
Mean (geometric)	<i>9.09nM</i>	<i>6.95nM</i>
(R-)-3-isobutyl GABA	157 207	78.4 64.2 107
Mean (geometric)	<i>180nM</i>	<i>81.5nM</i>

#### 5 Example 10

##### Filter binding assay of [<sup>3</sup>H]gabapentin binding to the recombinant polypeptide of SEQ ID No 9

- Assays were carried out at 21°C in a final volume of 250µl in 96-deep well plates. Assay
- 10 components were (all reagents were diluted in 10mM HEPES (pH 7.4 at 21°C)):
- 25µl compound to test
  - 200µl Polypeptide of SEQ ID No 9 (3µl protein diluted to 200µl)
  - 25µl radioligand ([<sup>3</sup>H]gabapentin (65Ci/mmol))
- 15 Plates were incubated at room temperature for 1h prior to filtering on to 96-well GF/B Unifilter plates pre-soaked in 0.3% polyethylenimine. Filters were washed with 3x1ml 50mM Tris-HCl (pH 7.4 at 4°C), and dried over-night. Scintillant (Microscint O, 50µl) was added and the plates counted using a Packard Top Count scintillation counter. Specific binding was ~98% of the 'total' value. In [<sup>3</sup>H]gabapentin saturation studies, the  $K_D$  (nM)
- 20 obtained was about 10.62.

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SEQUENCE LISTING**1- porcine nucleotide sequenc alpha2 delta-1**

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**2 - porcine nucleotide sequence**

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25 **3 - porcine nucleotide sequence**

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**4 - porcine nucleotide sequence**

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5

**5 - porcine amino acid sequence alpha2 delta-1**

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20 LLKPAVVGIKIDVNSWIENFTKTSIRDPCAGPVCDCRNSDVMDCVILDDGGFLLMANHDD  
YTNQIGRFFGEIDPSLMRHLVNI SVYAFNKSYDYQSVCEPGAAPKQGAGHRSAYVPSIADI  
LHIGWWATAAAWSILQQFLLSLTFPRLLEAVEMEDDDFTASLSKQSCITEQTQYFFDNDSK  
SFSGVLDCGNCSRI FHVEKLMNTNLIFIMVESKGTCPCDTRLLIQAEQTS DGPDPDCMVKQ  
PRYRKGPDVCFDNNALDYTD CGGVSGLNPSLWSIFGIQCVLLWLLSGSRHYQL

25

**6 - porcine amino acid sequence**

MAAGCLLALTTLTLFQSLLIGPSSQEPFPSAVTIKSWVDKMQEDLVTAKTASGVNQLVDIY  
EKYQDLYTVEPNNARQLVEIAARDIEKLLSNRSKALVRLALEAEKVQAAHQWREDFASNEV  
VYYNAKDDLDPEKNDSEPGSQRIKPVFIDANFGRQISYQHAAVHIPTDIYEGSTIVLNEL  
30 NWTALDEVFKKNREEDPSLLWQVFGSATGLARYYPASPWVDNSRTPNKIDLYDVRRRPWY  
IQGAASPKDMLILVDVSGSVSGLTLKLIRTSVSEMLETLSDDDFVNVASFNSNAQDVSCFQ  
HLVQANVRNKKVLKDAVNNITAKGITDYKKGFSFAFEQLLNYNVSRANCNKIIMLFTDGGE

ERAQEIFAKYNKDKKVRVFTFSVGQHNYDRGPIQWMACENKGYYYEIPSIGAIRINTQEYL  
DVLGRPMVLAGDKAKQVQWTNVYLDALGLVITGTLPVFNITGQENKTNLKNQLILGVM  
GVDVSLEDIKRLTPRFTLCPNGYYFAIDPNGYVLLHPNLQPKNPKSQEPVTLDFLDAELEN  
DIKVEIRNKMIDGESGEKTFRTL VKSQDERYIDKGNRTYTWTVPVNGTDYSLALVLPTYSFY  
5 YIKAKIEETITQARSKKGKMKDSETLKPDNFEESGYTFIAPRDYCNDLKISDNNTTEFLNLF  
NEFIDRKTPNNPSCNTDLINRVLLDAGFTNELVQNYWSKQKNIKGVKARFVVTGGITRVY  
PKEAGENWQENPETYEDSFYKRSLDNDNYVFTAPYFNKSGPGAYESGIMVSKAVEIYIQGK  
LLKPAVVGIKIDVNSWIENFTKTSIRDPCAGPVCDCKRNSDVMDCVILDDGGFLLMANHDD  
YTNQIGRFFGEIDPSLMRHLVNISVYAFNKSYDYQSVCEPGAAPKQGAGHRSAYVPSIADI  
10 LHIGWWATAAAWSILQQFLLSLTFPRLLLEAVEMEDDDFTASLSKQSCITEQTQYFFDNDSK  
SFSGVLDCGNCSRI FHVEKLMNTNLIFIMVESKGTCPCDTRL

**7 - porcine amino acid sequence**

MAAGCLLALTLTLFQSL LIGPSSQEPFPSAVTIKSWVDKMQEDLVT LAKTASGVNQLVDIY  
15 EKYQDLYTVEPNARQLVEIAARDIEKLLSNRSKALVRLALEAEKVQAAHQWREDFASNEV  
VYNAKDDLDPEKNDSEPGSQRIKPVFID DANFGRQISYQHAAVHIPTDIYEGSTIVLNEL  
NWT SALDEVFKKNREEDPSLLWQVFGSATGLARYYPASPWVDNSRTPNKIDLYDVRRRPWY  
IQGAASPKDMLILVDVSGSVSGLTLKLIRTSVSEMLETLSDDDFVNVASFNSNAQDVSCFQ  
HLVQANVRNKKVLKDAVN NITAKGITDYKKGFSFAFEQLLNYNVSRANCNKIIMLFTDGGE  
20 ERAQEIFAKYNKDKKVRVFTFSVGQHNYDRGPIQWMACENKGYYYEIPSIGAIRINTQEYL  
DVLGRPMVLAGDKAKQVQWTNVYLDALGLVITGTLPVFNITGQENKTNLKNQLILGVM  
GVDVSLEDIKRLTPRFTLCPNGYYFAIDPNGYVLLHPNLQPKNPKSQEPVTLDFLDAELEN  
DIKVEIRNKMIDGESGEKTFRTL VKSQDERYIDKGNRTYTWTVPVNGTDYSLALVLPTYSFY  
YIKAKIEETITQARSKKGKMKDSETLKPDNFEESGYTFIAPRDYCNDLKISDNNTTEFLNLF  
25 NEFIDRKTPNNPSCNTDLINRVLLDAGFTNELVQNYWSKQKNIKGVKARFVVTGGITRVY  
PKEAGENWQENPETYEDSFYKRSLDNDNYVFTAPYFNKSGPGAYESGIMVSKAVEIYIQGK  
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YTNQIGRFFGEIDPSLMRHLVNISVYAFNKSYDYQSVCEPGAAPKQGAGHRSAYVPSIADI  
LHIGWWATAAAWSILQQFLLSLTFPRLLLEAVEMEDDDFTASLSKQSCITEQTQYFFDNDSK  
30 SFSGVLDCGNCSRI FHVEKLMNTNLIFIMVESKGTCPCDTRLLIQAEQTS DGPDCDMVK

**8 - porcin amino acid sequence**

MAAGCLLALTLTLFQSLIGPSSQEPFPSAVTIKSWVDKMQEDLVTLAKTASGVNQLVDIY  
EKYQDLYTVEPNNAARQLVEIAARDIEKLLSNRSKALVRLALEAEKVQAAHQWREDFASNEV  
VYYNAKDDLDPEKNDSEPGSQRIKPVFIDANFGRQISYQHAAVHIPTDIYEGSTIVLNEL  
5 NWTALDEVFKKNREEDPSLLWQVFGSATGLARYYPASPWVDNSRTPNKIDLYDVRRRPWY  
IQGAASPKDMLILVDVSGSVSGLTLKLIRTSVSEMLETLSDDDFVNVASFNSNAQDVSCFQ  
HLVQANVRNKKVLKDAVNNITAKGITDYKKGFSFAFEQLLNYNVSRANCNKIIMLFTDGGGE  
ERAQEIFAQYNKDKKVRVFTFSVGQHNYDRGPIQWMACENKGYYYEIPSIGAIRINTQEYL  
DVLGRPMVLAGDKAKQVQWNTNVYLDALGLVITGTLPVFNITGQENKTNLKNQLILGVM  
10 GVDVSLEDIKRLTPRFTLCPNGYYFAIDPNGYVLLHPNLQPKNPKSQEPVTLDFLDAELEN  
DIKVEIRNKMIDGESGEKTFRTLKVSQDERYIDKGNRTYTWTVPVNGTDYSLALVLPTYSFY  
YIKAKIEETITQARSKKGKMKDSETLKPDNFEESGYTFIAPRDYCNLDKISDNNTFLNLF  
NEFIDRKT PNNPSCNTDLINRVLLDAGFTNELVQNYWSKQKNIGVKARFVVTGGITRVY  
PKEAGENWQENPETYEDSFYKRSNDNDNYFTAPYFNKSGPGAYESGIMVSKAVEIYIQGK  
15 LLKPAVVGKIDVNSWIENFTKTSIRDPCAGPVCDCKRNSDVMDCVILDDGGFLLMANHDD  
YTNQIGRFFGEIDPSLMRHLVNI SVYAFNKSYDYQSVCEPGAAPKQGAGHRSAYVPSIADI  
LHIGWWATAAAWSILQQFLSLTFPRLLLEAVEMEDDDFTASLSKQSCITEQTQYFFDNDK  
SFSGVLDCGNC SRIFHVEKLMNTNLIFIMVESKGTCPCDTRLLIQAEQTS DGPDPDCDMVKQ  
PRYRKGPDVCFDNNALDYTDCGGVS

20

**9 - porcine amino acid sequence**

MAAGCLLALTLTLFQSLIGPSSQEPFPSAVTIKSWVDKMQEDLVTLAKTASGVNQLVDIY  
EKYQDLYTVEPNNAARQLVEIAARDIEKLLSNRSKALVRLALEAEKVQAAHQWREDFASNEV  
VYYNAKDDLDPEKNDSEPGSQRIKPVFIDANFGRQISYQHAAVHIPTDIYEGSTIVLNEL  
25 NWTALDEVFKKNREEDPSLLWQVFGSATGLARYYPASPWVDNSRTPNKIDLYDVRRRPWY  
IQGAASPKDMLILVDVSGSVSGLTLKLIRTSVSEMLETLSDDDFVNVASFNSNAQDVSCFQ  
HLVQANVRNKKVLKDAVNNITAKGITDYKKGFSFAFEQLLNYNVSRANCNKIIMLFTDGGGE  
ERAQEIFAQYNKDKKVRVFTFSVGQHNYDRGPIQWMACENKGYYYEIPSIGAIRINTQEYL  
DVLGRPMVLAGDKAKQVQWNTNVYLDALGLVITGTLPVFNITGQENKTNLKNQLILGVM  
30 GVDVSLEDIKRLTPRFTLCPNGYYFAIDPNGYVLLHPNLQPKNPKSQEPVTLDFLDAELEN  
DIKVEIRNKMIDGESGEKTFRTLKVSQDERYIDKGNRTYTWTVPVNGTDYSLALVLPTYSFY  
YIKAKIEETITQARSKKGKMKDSETLKPDNFEESGYTFIAPRDYCNLDKISDNNTFLNLF

NEFIDRKTPNNPSCNTDLINRVLLDAGFTNELVQNYWSKQKNIKGVKARFVVTGGITRVY  
PKEAGENWQENPETYEDSFYKRSLDNDNYVFTAPYFNKSGPGAYESGIMVSKAVEIYIQGK  
LLKPAVVGIKIDVNSWIENFTKTSIRDPCAGPVCDCKRNSDVMDCVILDDGGFLLMANHDD  
YTNQIGRFFGEIDPSLMRHLVNI SVYAFNKS YDYQSVCEPGAAPKQGAGHRSAYVPSIADI  
5 LHIGWWATAAAWSILQQFLLSLTFPRLLEAVEMEDDDFTASLSKQSCITEQTQYFFDND SK  
SFSGVLDCGNCSRI FHVEKLMNTNLIFIMVESKGTCPCDTRLLIQAEQTS DGPDP CDMVKQ  
PRYRKGPDVCFDNNAL EDYTD CGGVSHHHHHH

**10 - human nucleotide sequence**

10 ATGGCTGCTGGCTGCCTGCTGGCCTTGACTCTGACACTTTTCCAATCTTTGCTCATCGGCC  
CCTCGTCGGAGGAGCCGTTCCCTTCGGCCGTC ACTATCAAATCATGGGTGGATAAGATGCA  
AGAAGACCTTGT CACACTGGCAAAAACAGCAAGTGGAGTCAATCAGCTTGT TGATATTTAT  
GAGAAATATCAAGATTTGTATACTGTGGAACCAAATAATGCACGCCAGCTGGTAGAAATTG  
CAGCCAGGGATATTGAGAACTTCTGAGCAACAGATCTAAAGCCCTGGTGAGCCTGGCATT  
15 GGAAGCGGAGAAAGTTCAAGCAGCTCACCAGTGGAGAGAAGATTTTGCAAGCAATGAAGTT  
GTCTACTACAATGCAAAGGATGATCTCGATCCTGAGAAAAATGACAGTGAGCCAGGCAGCC  
AGAGGATAAAACCTGTTTTTATTGAAGATGCTAATTTTGGACGACAAATATCTTATCAGCA  
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20 TGTGGCAGGTTTTTGGCAGTGCCACTGGCCTAGCTCGATATTATCCAGCTTCACCATGGGT  
TGATAATAGTAGAACTCAAATAAGATTGACCTTTATGATGTACGCAGAAGACCATGGTAC  
ATCCAAGGAGCTGCATCTCCTAAAGACATGCTTATTCTGGTGGATGTGAGTGGAAGTGTTA  
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25 CACCTTGTCCAAGCAAATGTAAGAAATAAAAAAGTGTTGAAAGACGCGGTGAATAATATCA  
CAGCCAAAGGAATTACAGATTATAAGAAGGGCTTTAGTTTTGCTTTTGAACAGCTGCTTAA  
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GAGAGAGCCCAGGAGATATTTAACAAATACAATAAAGATAAAAAAGTACGTGTATT CAGGT  
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30 AGGTTATTATTATGAAATTCCTTCCATTGGTGCAATAAGAATCAATACTCAGGAATATTTG  
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ATGTGTACCTGGATGCATTGGAAC TGGGACTTGTCATTACTGGAACCTTCCGGTCTTCAA

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5 GATATTAAAGTGGAGATTTCGAAATAAGATGATTGATGGGGAAAGTGGAGAAAAAACATTCA  
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10 AAGAGATTACTGCAATGACCTGAAAATATCGGATAATAACACTGAATTTCTTTTAAATTTTC  
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15 AAAGGAGCCTAGATAATGATAACTATGTTTTTCACTGCTCCCTACTTTAACAAAAGTGGACC  
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CGTAATGGATTGTGTGATTCTGGATGATGGTGGGTTTCTTCTGATGGCAAATCATGATGAT  
20 TATACTAATCAGATTGGAAGATTTTTTGGAGAGATTGATCCCAGCTTGATGAGACACCTGG  
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25 CCTGTCCAAGCAGAGCTGCATTACTGAACAAACCCAGTATTTCTTCGATAACGACAGTAAA  
TCATTCAGTGGTGTATTAGACTGTGGAACTGTTCCAGAATCTTTCATGGAGAAAAGCTTA  
TGAACACCAACTTAATATTCATAATGGTTGAGAGCAAAGGGACATGTCCATGTGACACACG  
ACTGC

30 11 - human nucleotide sequence

ATGGCTGCTGGCTGCCTGCTGGCCTTGACTCTGACACTTTTCCAATCTTTGCTCATCGGCC  
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5 GTCTACTACAATGCAAAGGATGATCTCGATCCTGAGAAAAATGACAGTGAGCCAGGCAGCC  
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10 TGATAATAGTAGAACTCCAAATAAGATTGACCTTTATGATGTACGCAGAAGACCATGGTAC  
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TGATGATTTTCGTGAATGTAGCTTCATTTAACAGCAATGCTCAGGATGTAAGCTGTTTTCAG  
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15 CAGCCAAAGGAATTACAGATTATAAGAAGGGCTTTAGTTTTGCTTTTGAACAGCTGCTTAA  
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AGGTTATTATTATGAAATTCCTTCCATTGGTGCAATAAGAATCAATACTCAGGAATATTTG  
20 GATGTTTTGGGAAGACCAATGGTTTTAGCAGGAGACAAAGCTAAGCAAGTCCAATGGACAA  
ATGTGTACCTGGATGCATTGGAAGTGGGACTTGTCATTACTGGAAGTCTTCCGGTCTTCAA  
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ATGGGTATTACTTTGCAATCGATCCTAATGGTTATGTTTTATTACATCCAAATCTTCAGCC  
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30 AGGATTCGGAAACCCTGAAGCCAGATAATTTGAAGAATCTGGCTATACATTCATAGCACC  
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AACGAGTTTATTGATAGAAAACTCCAAACAACCCATCATGTAACGCGGATTTGATTAATA

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5 TGGTGCCTATGAATCGGGCATTATGGTAAGCAAAGCTGTAGAAATATATATTCAAGGGAAA  
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10 TTAATATATCAGTTTATGCTTTTAAACAAATCTTATGATTATCAGTCAGTATGTGAGCCCGG  
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CCTGTCCAAGCAGAGCTGCATTACTGAACAAACCCAGTATTTCTTCGATAACGACAGTAAA  
15 TCATTCAGTGGTGTATTAGACTGTGGAACTGTTCCAGAATCTTTCATGGAGAAAAGCTTA  
TGAACACCAACTTAATATTCATAATGGTTGAGAGCAAAGGGACATGTCCATGTGACACACG  
ACTGCTCATACAAGCGGAGCAGACTTCTGACGGTCCAAATCCTTGTGACATGGTTAAGC

## 12 - human nucleotide sequence

20 ATGGCTGCTGGCTGCCTGCTGGCCTTGACTCTGACACTTTTCCAATCTTTGCTCATCGGCC  
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CAGCCAGGGATATTGAGAACTTCTGAGCAACAGATCTAAAGCCCTGGTGGCCTGGCATT  
25 GGAAGCGGAGAAAGTTCAAGCAGCTCACCAGTGGAGAGAAGATTTTGCAAGCAATGAAGTT  
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15 GATATTAAAGTGGAGATTGCAATAAGATGATTGATGGGGAAAGTGGAGAAAAACATTCA  
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20 AAGAGATTACTGCAATGACCTGAAAATATCGGATAATAACACTGAATTTCTTTTAAATTTT  
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30 TATACTAATCAGATTGGAAGATTTTTTGGAGAGATTGATCCCAGCTTGATGAGACACCTGG  
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TTACAAATTGGCTGGTGGGCCACTGCTGCTGCCTGGTCTATTCTACAGCAGTTTCTCTTGA  
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CCTGTCCAAGCAGAGCTGCATTACTGAACAAACCCAGTATTTCTTCGATAACGACAGTAAA  
TCATTCAGTGGTGTATTAGACTGTGGAACTGTTCCAGAATCTTTCATGGAGAAAAGCTTA  
5 TGAACACCAACTTAATATTCATAATGGTTGAGAGCAAAGGGACATGTCCATGTGACACAG  
ACTGCTCATACAAGCGGAGCAGACTTCTGACGGTCCAAATCCTTGTGACATGGTTAAGCAA  
CCTAGATACCGAAAAGGGCCTGATGTCTGCTTTGATAACAATGTCTTGGAGGATTATACTG  
ACTGTGGTGGTGTCTTCTG

10 13 - human amino acid sequence

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EKYQDLYTVEPNNAARQLVEIAARDIEKLLSNRSKALVSLALEAEKVQAAHQWREDFASNEV  
VYYNAKDDLDPEKNDSEPGSQRIKPVFIEDANFGRQISYQHAAVHIPTDIYEGSTIVLNEL  
NWTSALEDEVFKKNREEDPSLLWQVFGSATGLARYYPASPWVNSRTPNKIDLYDVRRRPWY  
15 IQGAASPKDMLILVDVSGSVSGLTLKLIRTSVSEMLETLSDDDFVNVASFNSNAQDVSCFQ  
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14 - human amino acid sequence

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15

**15 - human amino acid sequence**

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**17 - human alpha2 delta-1 nucleic acid sequence**

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**18 - nucleotide sequence**

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**25 19 - nucleotide sequence**

CTGAGATTTGGGGTTCTTTGG

**20 - nucleotide sequence**

TCGCCACCATGGCTGCTGGCTGCCTGCTG

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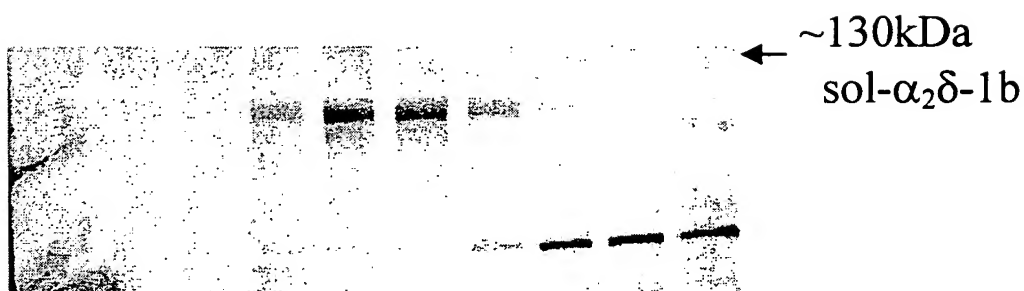
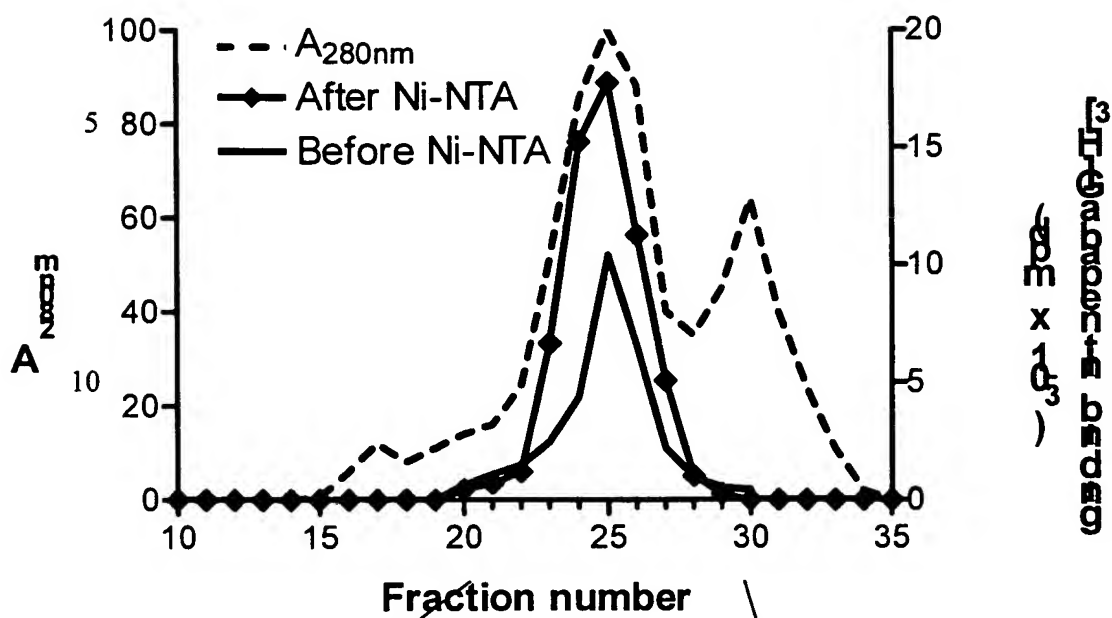
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**CLAIMS:**

1. A method for the screening of ligands which bind a cerebral cortical voltage-dependent calcium channel  $\alpha_2\delta$ -1 subunit, said method comprising the steps of:
- 5       - contacting a secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide with:
- a ligand of interest; and
- a labelled compound which binds the  $\alpha_2\delta$ -1 subunit; and
- 10       - measuring the level of binding of the labelled compound to the  $\alpha_2\delta$ -1 subunit.
2. A method according to claim 1, wherein said contacting and said binding is in a well of a flashplate.
- 15   3. A method according to claim 1, wherein said secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide is selected from the group consisting of SEQ ID NO: 6, 7, 8, 9, 13, 14 and 15.
4. A method according to claim 1, wherein said secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide is selected from the group consisting of SEQ ID NO: 9 and 15.
- 20   5. A method according to claim 1, wherein said secreted soluble recombinant calcium channel  $\alpha_2\delta$ -1 subunit polypeptide is SEQ ID NO: 9.

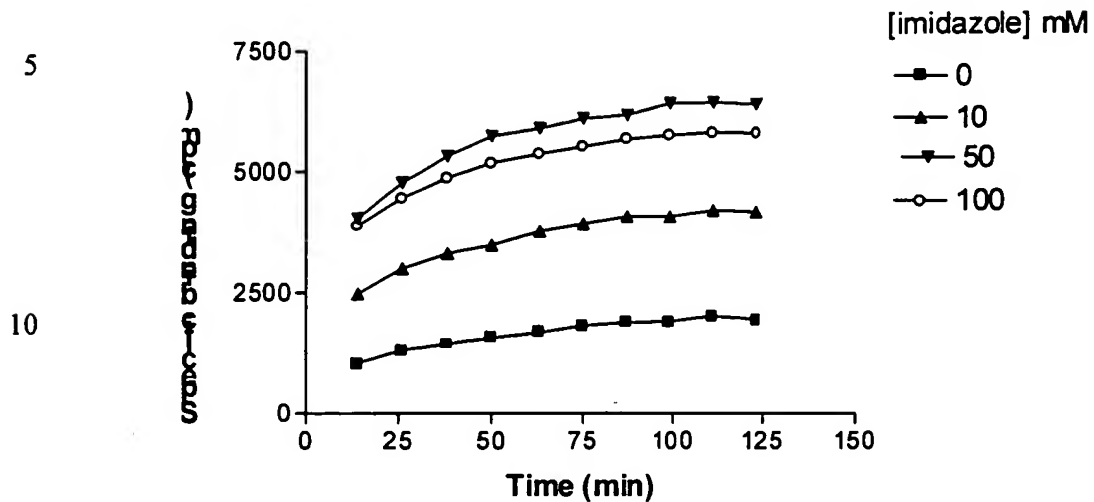
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**FIGURE 1**

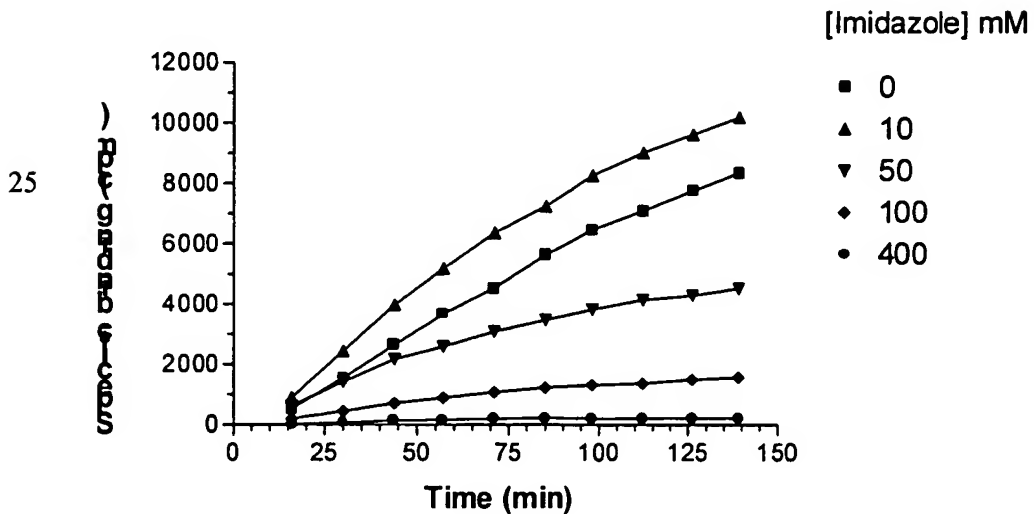


**FIGURE 2**

SPA assay of [ $^3\text{H}$ ]gabapentin (18.4nM) binding to s- $\alpha_2\delta$ -1b-6His (20 $\mu\text{l}$ ). Optimisation of Imidazole concentration in the assay.

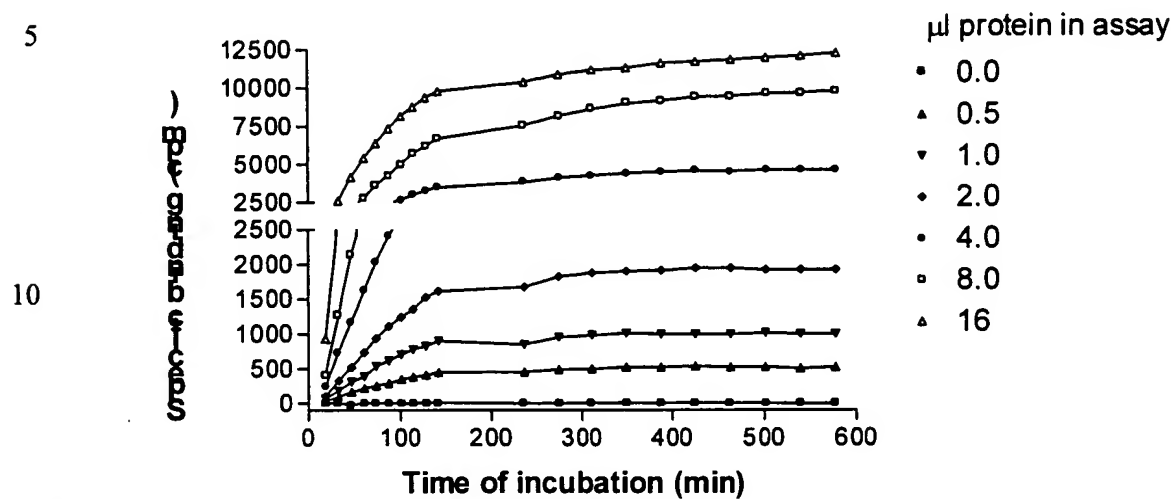
**FIGURE 3**

Flashplate assay of [ $^3\text{H}$ ]gabapentin (14nM) binding to s- $\alpha_2\delta$ -1b-6His (10 $\mu\text{l}$ ). Optimisation of Imidazole concentration in the assay.



**FIGURE 4**

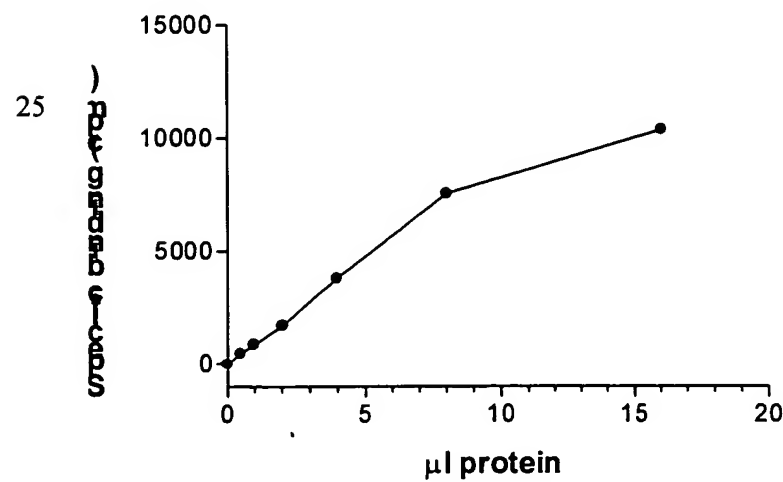
Flashplate time course of [<sup>3</sup>H]gabapentin (13nM) binding to various concentrations of s- $\alpha_2\delta$ -1b-6His.



10mM imidazole in assay  
15

**FIGURE 5**

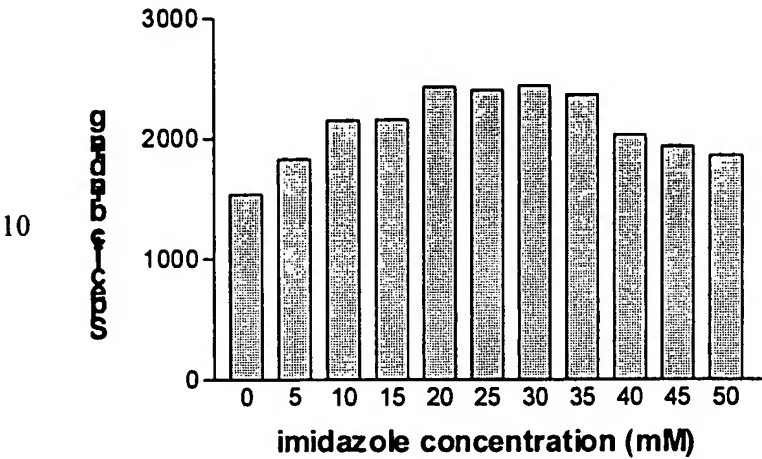
Determination of s- $\alpha_2\delta$ -1b-6His capacity of flashplate assay. Counted after 3hour incubation



**FIGURE 6**

Determination of the optimum imidazole concentration required to maximize the [<sup>3</sup>H]gabapentin (13nM) binding window using a constant amount of purified s-α<sub>2</sub>δ-1b-6His (2μl). Assayed after 3hour incubation.

5



15

**FIGURE 7**

Flashplate assay of [<sup>3</sup>H]gabapentin saturation binding to purified s-α<sub>2</sub>δ-1b-6His. Assayed after three hour incubation (see table 1 for details).

20

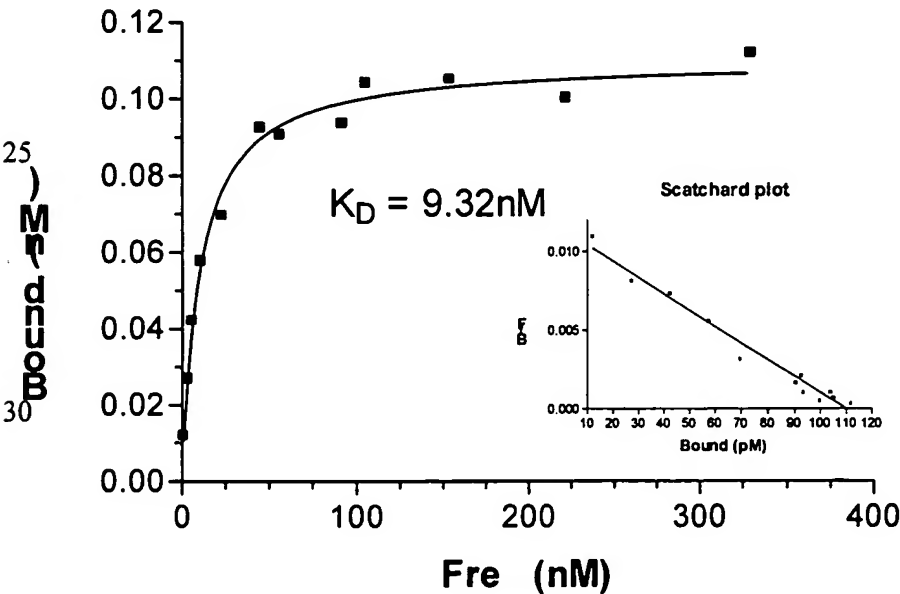
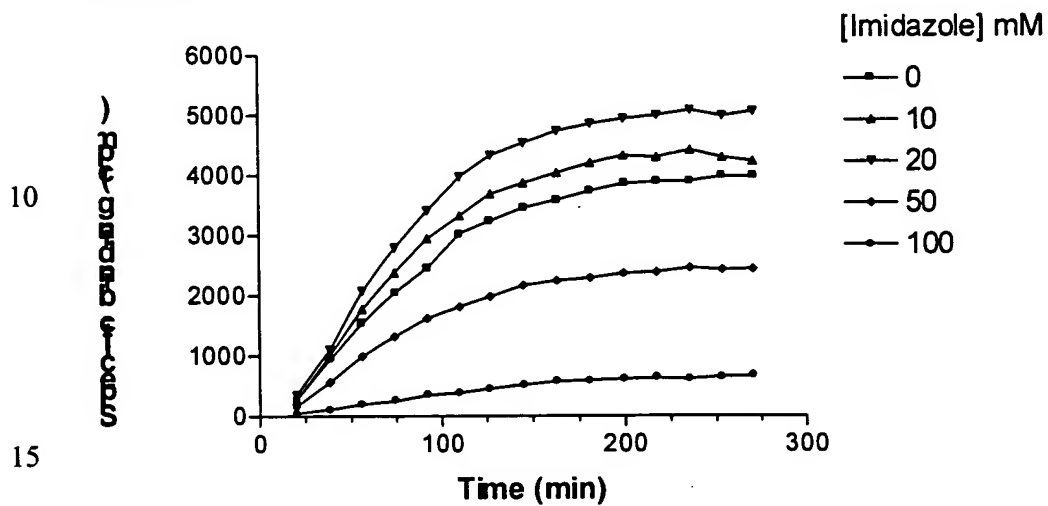
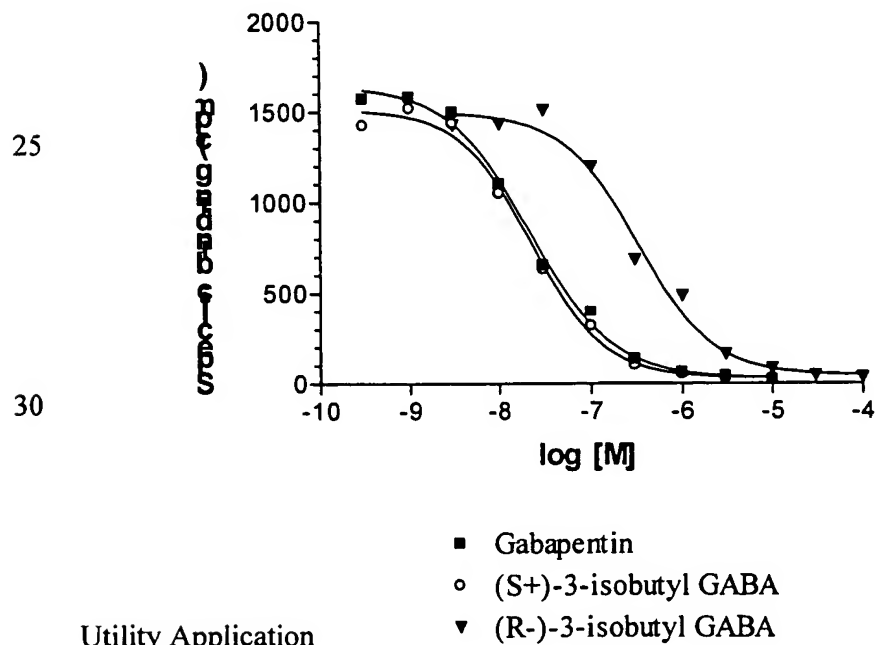


Figure 8

Flashplate time course optimisation of Imidazole concentration required to maximize the  $[^3\text{H}]$ Leucine (10.1nM) binding window to s- $\alpha_2\delta$ -1b-6His. Assayed after three hour incubation.

**FIGURE 9**

Competition curves of three compounds in the flashplate assay format (see table 2 for details). Assayed after 3 hour incubation.



5

**ABSTRACT**

10

**Method for the screening of  $\alpha_2\delta$ -1 subunit binding ligands**

15 A method for the screening of ligands which bind to soluble  $\alpha_2\delta$ -1 subtype polypeptides.

IN THE UNITED STATES PATENT & TRADEMARK OFFICE

APPLICANT : Francois Bertelli et al

EXAMINER :

SERIAL NO. :

ART UNIT :

FILED : Herewith

PAPER NO. :

FOR : Method For The Screening of Alpha2Delta Subunit Binding Ligands

**Request for Transfer of Computer Readable Form Under 37 CFR 1.821(e)**

March 6, 2002


Commissioner for Patent  
Washington, D.C. 20231

Dear Sir:

The paper copy of the Sequence Listing in this application, is identical to the computer readable copy of the Sequence Listing filed in Application No. 09/397,549, Filed September 16, 1999. In accordance with 37CFR 1.821(e), please use the only filed computer readable form filed in that application as the computer readable form for the instant application. It is understood that the Patent and Trademark Office will make necessary change in the application number and filing date for the instant application. A paper copy of the Sequence Listing is included in the originally filed specification of the instant application.

Respectfully submitted,

Dated: March 6, 2002



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EXPRESS MAIL NO. EJ054450643US  
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# SEQUENCE LISTING

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Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu	450	455	460
Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Asn	Glu	Asn	Lys	Thr	Asn	Leu	Lys	465	470	475
Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp	485	490	495
Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr	500	505	510
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln	515	520	525
Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp	530	535	540
Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile	545	550	555
Asp	Gly	Glu	Ser	Gly	Glu	Lys	Thr	Phe	Arg	Thr	Leu	Val	Lys	Ser	Gln	565	570	575
Asp	Glu	Arg	Tyr	Ile	Asp	Lys	Gly	Asn	Arg	Thr	Tyr	Thr	Trp	Thr	Pro	580	585	590
Val	Asn	Gly	Thr	Asp	Tyr	Ser	Leu	Ala	Leu	Val	Leu	Pro	Thr	Tyr	Ser	595	600	605
Phe	Tyr	Tyr	Ile	Lys	Ala	Lys	Ile	Glu	Glu	Thr	Ile	Thr	Gln	Ala	Arg	610	615	620
Ser	Lys	Lys	Gly	Lys	Met	Lys	Asp	Ser	Glu	Thr	Leu	Lys	Pro	Asp	Asn	625	630	635
Phe	Glu	Glu	Ser	Gly	Tyr	Thr	Phe	Ile	Ala	Pro	Arg	Asp	Tyr	Cys	Asn	645	650	655

Asp	Leu	Lys	Ile	Ser	Asp	Asn	Asn	Thr	Glu	Phe	Leu	Leu	Asn	Phe	Asn		
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Glu	Phe	Ile	Asp	Arg	Lys	Thr	Pro	Asn	Asn	Pro	Ser	Cys	Asn	Thr	Asp		
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Leu	Ile	Asn	Arg	Val	Leu	Leu	Asp	Ala	Gly	Phe	Thr	Asn	Glu	Leu	Val		
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Gln	Asn	Tyr	Trp	Ser	Lys	Gln	Lys	Asn	Ile	Lys	Gly	Val	Lys	Ala	Arg		
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Lys	Arg	Ser	Leu	Asp	Asn	Asp	Asn	Tyr	Val	Phe	Thr	Ala	Pro	Tyr	Phe		
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Ser	Ile	Arg	Asp	Pro	Cys	Ala	Gly	Pro	Val	Cys	Asp	Cys	Lys	Arg	Asn		
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Ser	Asp	Val	Met	Asp	Cys	Val	Ile	Leu	Asp	Asp	Gly	Gly	Phe	Leu	Leu		
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Met	Ala	Asn	His	Asp	Asp	Tyr	Thr	Asn	Gln	Ile	Gly	Arg	Phe	Phe	Gly		
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Glu	Ile	Asp	Pro	Ser	Leu	Met	Arg	His	Leu	Val	Asn	Ile	Ser	Val	Tyr		
865					870					875					880		
Ala	Phe	Asn	Lys	Ser	Tyr	Asp	Tyr	Gln	Ser	Val	Cys	Glu	Pro	Gly	Ala		
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Ala	Pro	Lys	Gln	Gly	Ala	Gly	His	Arg	Ser	Ala	Tyr	Val	Pro	Ser	Ile		
		900						905					910				
Ala	Asp	Ile	Leu	His	Ile	Gly	Trp	Trp	Ala	Thr	Ala	Ala	Ala	Trp	Ser		
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Ile	Leu	Gln	Gln	Phe	Leu	Leu	Ser	Leu	Thr	Phe	Pro	Arg	Leu	Leu	Glu		
	930					935					940						
Ala	Val	Glu	Met	Glu	Asp	Asp	Asp	Phe	Thr	Ala	Ser	Leu	Ser	Lys	Gln		
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Ser Cys Ile Thr Glu Gln Thr Gln Tyr Phe Phe Asp Asn Asp Ser Lys  
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 Ser Phe Ser Gly Val Leu Asp Cys Gly Asn Cys Ser Arg Ile Phe His  
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 Val Glu Lys Leu Met Asn Thr Asn Leu Ile Phe Ile Met Val Glu Ser  
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 Lys Gly Thr Cys Pro Cys Asp Thr Arg Leu Leu Ile Gln Ala Glu Gln  
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 Thr Ser Asp Gly Pro Asp Pro Cys Asp Met Val Lys Gln Pro Arg Tyr  
 1025 1030 1035 1040  
 Arg Lys Gly Pro Asp Val Cys Phe Asp Asn Asn Ala Leu Glu Asp Tyr  
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 Thr Asp Cys Gly Gly Val Ser Gly Leu Asn Pro Ser Leu Trp Ser Ile  
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 Ile Lys Ser Trp Val Asp Lys Met Gln Glu Asp Leu Val Thr Leu Ala  
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 Lys Thr Ala Ser Gly Val Asn Gln Leu Val Asp Ile Tyr Glu Lys Tyr  
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 Gln Asp Leu Tyr Thr Val Glu Pro Asn Asn Ala Arg Gln Leu Val Glu  
 65 70 75 80  
 Ile Ala Ala Arg Asp Ile Glu Lys Leu Leu Ser Asn Arg Ser Lys Ala  
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 Leu Val Arg Leu Ala Leu Glu Ala Glu Lys Val Gln Ala Ala His Gln  
 100 105 110  
 Trp Arg Glu Asp Phe Ala Ser Asn Glu Val Val Tyr Tyr Asn Ala Lys

115					120					125					
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Tyr	Gln	His	Ala	Ala	Val	His	Ile	Pro	Thr	Asp	Ile	Tyr	Glu	Gly	Ser
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Thr	Ile	Val	Leu	Asn	Glu	Leu	Asn	Trp	Thr	Ser	Ala	Leu	Asp	Glu	Val
			180					185					190		
Phe	Lys	Lys	Asn	Arg	Glu	Glu	Asp	Pro	Ser	Leu	Leu	Trp	Gln	Val	Phe
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Gly	Ser	Ala	Thr	Gly	Leu	Ala	Arg	Tyr	Tyr	Pro	Ala	Ser	Pro	Trp	Val
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Asp	Asn	Ser	Arg	Thr	Pro	Asn	Lys	Ile	Asp	Leu	Tyr	Asp	Val	Arg	Arg
225						230					235				240
Arg	Pro	Trp	Tyr	Ile	Gln	Gly	Ala	Ala	Ser	Pro	Lys	Asp	Met	Leu	Ile
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Leu	Val	Asp	Val	Ser	Gly	Ser	Val	Ser	Gly	Leu	Thr	Leu	Lys	Leu	Ile
			260					265					270		
Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe
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Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe
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Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp
305						310					315				320
Ala	Val	Asn	Asn	Ile	Thr	Ala	Lys	Gly	Ile	Thr	Asp	Tyr	Lys	Lys	Gly
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Phe	Ser	Phe	Ala	Phe	Glu	Gln	Leu	Leu	Asn	Tyr	Asn	Val	Ser	Arg	Ala
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Ala	Gln	Glu	Ile	Phe	Ala	Lys	Tyr	Asn	Lys	Asp	Lys	Lys	Val	Arg	Val
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385						390					395				400
Trp	Met	Ala	Cys	Glu	Asn	Lys	Gly	Tyr	Tyr	Tyr	Glu	Ile	Pro	Ser	Ile
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Gly	Ala	Ile	Arg	Ile	Asn	Thr	Gln	Glu	Tyr	Leu	Asp	Val	Leu	Gly	Arg



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Pro	Met	Val	Leu	Ala	Gly	Asp	Lys	Ala	Lys	Gln	Val	Gln	Trp	Thr	Asn
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Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu
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Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp
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Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr
			500						505			510			
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln
			515						520			525			
Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp
530						535						540			
Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile
545						550						555			
Asp	Gly	Glu	Ser	Gly	Glu	Lys	Thr	Phe	Arg	Thr	Leu	Val	Lys	Ser	Gln
			565						570			575			
Asp	Glu	Arg	Tyr	Ile	Asp	Lys	Gly	Asn	Arg	Thr	Tyr	Thr	Trp	Thr	Pro
			580						585			590			
Val	Asn	Gly	Thr	Asp	Tyr	Ser	Leu	Ala	Leu	Val	Leu	Pro	Thr	Tyr	Ser
			595						600			605			
Phe	Tyr	Tyr	Ile	Lys	Ala	Lys	Ile	Glu	Glu	Thr	Ile	Thr	Gln	Ala	Arg
610						615						620			
Ser	Lys	Lys	Gly	Lys	Met	Lys	Asp	Ser	Glu	Thr	Leu	Lys	Pro	Asp	Asn
625						630						635			
Phe	Glu	Glu	Ser	Gly	Tyr	Thr	Phe	Ile	Ala	Pro	Arg	Asp	Tyr	Cys	Asn
			645						650			655			
Asp	Leu	Lys	Ile	Ser	Asp	Asn	Asn	Thr	Glu	Phe	Leu	Leu	Asn	Phe	Asn
			660						665			670			
Glu	Phe	Ile	Asp	Arg	Lys	Thr	Pro	Asn	Asn	Pro	Ser	Cys	Asn	Thr	Asp
675						680						685			
Leu	Ile	Asn	Arg	Val	Leu	Leu	Asp	Ala	Gly	Phe	Thr	Asn	Glu	Leu	Val
690						695						700			
Gln	Asn	Tyr	Trp	Ser	Lys	Gln	Lys	Asn	Ile	Lys	Gly	Val	Lys	Ala	Arg
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 Ile Lys Ser Trp Val Asp Lys Met Gln Glu Asp Leu Val Thr Leu Ala  
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 Lys Thr Ala Ser Gly Val Asn Gln Leu Val Asp Ile Tyr Glu Lys Tyr  
                   50                  55                  60  
 Gln Asp Leu Tyr Thr Val Glu Pro Asn Asn Ala Arg Gln Leu Val Glu  
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 Ile Ala Ala Arg Asp Ile Glu Lys Leu Leu Ser Asn Arg Ser Lys Ala  
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 Leu Val Arg Leu Ala Leu Glu Ala Glu Lys Val Gln Ala Ala His Gln  
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 Trp Arg Glu Asp Phe Ala Ser Asn Glu Val Val Tyr Tyr Asn Ala Lys  
                   115                  120                  125  
 Asp Asp Leu Asp Pro Glu Lys Asn Asp Ser Glu Pro Gly Ser Gln Arg  
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 Ile Lys Pro Val Phe Ile Asp Asp Ala Asn Phe Gly Arg Gln Ile Ser  
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 Tyr Gln His Ala Ala Val His Ile Pro Thr Asp Ile Tyr Glu Gly Ser  
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 Thr Ile Val Leu Asn Glu Leu Asn Trp Thr Ser Ala Leu Asp Glu Val  
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 Phe Lys Lys Asn Arg Glu Glu Asp Pro Ser Leu Leu Trp Gln Val Phe  
                   195                  200                  205  
 Gly Ser Ala Thr Gly Leu Ala Arg Tyr Tyr Pro Ala Ser Pro Trp Val  
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 Asp Asn Ser Arg Thr Pro Asn Lys Ile Asp Leu Tyr Asp Val Arg Arg  
                   225                  230                  235                  240  
 Arg Pro Trp Tyr Ile Gln Gly Ala Ala Ser Pro Lys Asp Met Leu Ile  
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 Leu Val Asp Val Ser Gly Ser Val Ser Gly Leu Thr Leu Lys Leu Ile  
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Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe	275	280	285
Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe	290	295	300
Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp	305	310	315
Ala	Val	Asn	Asn	Ile	Thr	Ala	Lys	Gly	Ile	Thr	Asp	Tyr	Lys	Lys	Gly	325	330	335
Phe	Ser	Phe	Ala	Phe	Glu	Gln	Leu	Leu	Asn	Tyr	Asn	Val	Ser	Arg	Ala	340	345	350
Asn	Cys	Asn	Lys	Ile	Ile	Met	Leu	Phe	Thr	Asp	Gly	Gly	Glu	Glu	Arg	355	360	365
Ala	Gln	Glu	Ile	Phe	Ala	Lys	Tyr	Asn	Lys	Asp	Lys	Lys	Val	Arg	Val	370	375	380
Phe	Thr	Phe	Ser	Val	Gly	Gln	His	Asn	Tyr	Asp	Arg	Gly	Pro	Ile	Gln	385	390	395
Trp	Met	Ala	Cys	Glu	Asn	Lys	Gly	Tyr	Tyr	Tyr	Glu	Ile	Pro	Ser	Ile	405	410	415
Gly	Ala	Ile	Arg	Ile	Asn	Thr	Gln	Glu	Tyr	Leu	Asp	Val	Leu	Gly	Arg	420	425	430
Pro	Met	Val	Leu	Ala	Gly	Asp	Lys	Ala	Lys	Gln	Val	Gln	Trp	Thr	Asn	435	440	445
Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu	450	455	460
Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Asn	Glu	Asn	Lys	Thr	Asn	Leu	Lys	465	470	475
Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp	485	490	495
Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr	500	505	510
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln	515	520	525
Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp	530	535	540
Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile	545	550	555
Asp	Gly	Glu	Ser	Gly	Glu	Lys	Thr	Phe	Arg	Thr	Leu	Val	Lys	Ser	Gln	565	570	575

Asp	Glu	Arg	Tyr	Ile	Asp	Lys	Gly	Asn	Arg	Thr	Tyr	Thr	Trp	Thr	Pro		
			580					585					590				
Val	Asn	Gly	Thr	Asp	Tyr	Ser	Leu	Ala	Leu	Val	Leu	Pro	Thr	Tyr	Ser		
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Phe	Tyr	Tyr	Ile	Lys	Ala	Lys	Ile	Glu	Glu	Thr	Ile	Thr	Gln	Ala	Arg		
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Ser	Lys	Lys	Gly	Lys	Met	Lys	Asp	Ser	Glu	Thr	Leu	Lys	Pro	Asp	Asn		
	625				630					635					640		
Phe	Glu	Glu	Ser	Gly	Tyr	Thr	Phe	Ile	Ala	Pro	Arg	Asp	Tyr	Cys	Asn		
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Asp	Leu	Lys	Ile	Ser	Asp	Asn	Asn	Thr	Glu	Phe	Leu	Leu	Asn	Phe	Asn		
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Glu	Phe	Ile	Asp	Arg	Lys	Thr	Pro	Asn	Asn	Pro	Ser	Cys	Asn	Thr	Asp		
		675					680					685					
Leu	Ile	Asn	Arg	Val	Leu	Leu	Asp	Ala	Gly	Phe	Thr	Asn	Glu	Leu	Val		
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Gln	Asn	Tyr	Trp	Ser	Lys	Gln	Lys	Asn	Ile	Lys	Gly	Val	Lys	Ala	Arg		
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Phe	Val	Val	Thr	Asp	Gly	Gly	Ile	Thr	Arg	Val	Tyr	Pro	Lys	Glu	Ala		
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Gly	Glu	Asn	Trp	Gln	Glu	Asn	Pro	Glu	Thr	Tyr	Glu	Asp	Ser	Phe	Tyr		
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Lys	Arg	Ser	Leu	Asp	Asn	Asp	Asn	Tyr	Val	Phe	Thr	Ala	Pro	Tyr	Phe		
		755					760					765					
Asn	Lys	Ser	Gly	Pro	Gly	Ala	Tyr	Glu	Ser	Gly	Ile	Met	Val	Ser	Lys		
	770					775					780						
Ala	Val	Glu	Ile	Tyr	Ile	Gln	Gly	Lys	Leu	Leu	Lys	Pro	Ala	Val	Val		
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Gly	Ile	Lys	Ile	Asp	Val	Asn	Ser	Trp	Ile	Glu	Asn	Phe	Thr	Lys	Thr		
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Ser	Ile	Arg	Asp	Pro	Cys	Ala	Gly	Pro	Val	Cys	Asp	Cys	Lys	Arg	Asn		
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Ser	Asp	Val	Met	Asp	Cys	Val	Ile	Leu	Asp	Asp	Gly	Gly	Phe	Leu	Leu		
			835				840					845					
Met	Ala	Asn	His	Asp	Asp	Tyr	Thr	Asn	Gln	Ile	Gly	Arg	Phe	Phe	Gly		
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Glu	Ile	Asp	Pro	Ser	Leu	Met	Arg	His	Leu	Val	Asn	Ile	Ser	Val	Tyr		
	865				870					875					880		



Leu Val Arg Leu Ala Leu Glu Ala Glu Lys Val Gln Ala Ala His Gln  
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 Trp Arg Glu Asp Phe Ala Ser Asn Glu Val Val Tyr Tyr Asn Ala Lys  
 115 120 125  
 Asp Asp Leu Asp Pro Glu Lys Asn Asp Ser Glu Pro Gly Ser Gln Arg  
 130 135 140  
 Ile Lys Pro Val Phe Ile Asp Asp Ala Asn Phe Gly Arg Gln Ile Ser  
 145 150 155 160  
 Tyr Gln His Ala Ala Val His Ile Pro Thr Asp Ile Tyr Glu Gly Ser  
 165 170 175  
 Thr Ile Val Leu Asn Glu Leu Asn Trp Thr Ser Ala Leu Asp Glu Val  
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 Phe Lys Lys Asn Arg Glu Glu Asp Pro Ser Leu Leu Trp Gln Val Phe  
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 Gly Ser Ala Thr Gly Leu Ala Arg Tyr Tyr Pro Ala Ser Pro Trp Val  
 210 215 220  
 Asp Asn Ser Arg Thr Pro Asn Lys Ile Asp Leu Tyr Asp Val Arg Arg  
 225 230 235 240  
 Arg Pro Trp Tyr Ile Gln Gly Ala Ala Ser Pro Lys Asp Met Leu Ile  
 245 250 255  
 Leu Val Asp Val Ser Gly Ser Val Ser Gly Leu Thr Leu Lys Leu Ile  
 260 265 270  
 Arg Thr Ser Val Ser Glu Met Leu Glu Thr Leu Ser Asp Asp Asp Phe  
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 Val Asn Val Ala Ser Phe Asn Ser Asn Ala Gln Asp Val Ser Cys Phe  
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 Ala Val Asn Asn Ile Thr Ala Lys Gly Ile Thr Asp Tyr Lys Lys Gly  
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 Phe Ser Phe Ala Phe Glu Gln Leu Leu Asn Tyr Asn Val Ser Arg Ala  
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 Asn Cys Asn Lys Ile Ile Met Leu Phe Thr Asp Gly Gly Glu Glu Arg  
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 Ala Gln Glu Ile Phe Ala Lys Tyr Asn Lys Asp Lys Lys Val Arg Val  
 370 375 380  
 Phe Thr Phe Ser Val Gly Gln His Asn Tyr Asp Arg Gly Pro Ile Gln  
 385 390 395 400

Trp	Met	Ala	Cys	Glu	Asn	Lys	Gly	Tyr	Tyr	Tyr	Glu	Ile	Pro	Ser	Ile	
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Gly	Ala	Ile	Arg	Ile	Asn	Thr	Gln	Glu	Tyr	Leu	Asp	Val	Leu	Gly	Arg	
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Pro	Met	Val	Leu	Ala	Gly	Asp	Lys	Ala	Lys	Gln	Val	Gln	Trp	Thr	Asn	
		435					440					445				
Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu	
	450					455					460					
Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Asn	Glu	Asn	Lys	Thr	Asn	Leu	Lys	
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Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp	
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Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr	
			500					505					510			
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln	
	515						520					525				
Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp	
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Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile	
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attgacaaa	gaaacaggac	atacacatgg	acacctgtca	atggcacaga	ttacagtttg	1800
gccttggtat	taccaacctta	cagttttttac	tatataaaa	ccaaactaga	agagacaata	1860
actcaggcca	gatcaaaaaa	gggcaaaatg	aaggattcgg	aaaccctgaa	gccagataat	1920
tttgaagaat	ctggctatac	attcatagca	ccaagagatt	actgcaatga	cctgaaaata	1980
tcggataata	acactgaatt	tcttttaaat	ttcaacgagt	ttattgatag	aaaaactcca	2040
aacaacccat	catgtaacgc	ggatttgatt	aatagagtct	tgcttgatgc	aggctttaca	2100
aatgaacttg	tccaaaatta	ctggagtaag	cagaaaaata	tcaagggagt	gaaagcacga	2160
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caagaaaacc	cagagacata	tgaggacagc	ttctataaaa	ggagcctaga	taatgataac	2280
tatgtttttca	ctgctcccta	ctttaacaaa	agtggacctg	gtgcctatga	atcgggcatt	2340
atggtaaagca	aagctgtaga	aatatatatt	caagggaac	ttcttaaaacc	tgcatgtgtt	2400
ggaattaaaa	ttgatgtaaa	ttcctggata	gagaatttca	ccaaaacctc	aatcagagat	2460
ccgtgtgctg	gtccagtttg	tgactgcaaa	agaaacagtg	acgtaatgga	ttgtgtgatt	2520
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agattttttg	gagagattga	tcccagcttg	atgagacacc	tggttaatat	atcagtttat	2640
gcttttaaca	aatcttatga	ttatcagtca	gtatgtgagc	ccggtgctgc	acaaaaacaa	2700
ggagcaggac	atcgctcagc	atatgtgcca	tcagtagcag	acatattaca	aattggctgg	2760
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cgactccttg	aggcagttga	gatggaggat	gatgacttca	cggcctccct	gtccaagcag	2880
agctgcatta	ctgaacaaac	ccagtatttc	ttcgataacg	acagtaaate	attcagtggt	2940
gtattagact	gtggaaactg	ttccagaatc	tttcatggag	aaaagcttat	gaacaccaac	3000
ttaatatcca	taatggttga	gagcaaaggg	acatgtccat	gtgacacacg	actgctcata	3060
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 <211> 3190  
 <212> DNA  
 <213> Homo sapiens

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 caagaagacc ttgtcacact ggcaaaaaca gcaagtggag tcaatcagct tgttgatatt 180  
 tatgagaaat atcaagattt gtatactgtg gaaccaaata atgcacgcca gctggtagaa 240  
 attgcagcca gggatattga gaaactttct agcaacagat ctaaagccct ggtgagcctg 300  
 gcattggaag cggagaaaagt tcaagcagct caccagtga gagaagattt tgcaagcaat 360  
 gaagttgtct actacaatgc aaaggatgat ctcgatccct agaaaaatga cagtgaacca 420  
 ggcagccaga ggataaaacc tgttttcatt gaagatgcta attttggacg acaaatatct 480  
 tatcagcagc cagcagtcca tttcctact gacatctatg agggctcaac aatttgttta 540  
 aatgaactca actggacaag tgccttagat gaagttttca aaaagaatcg cgaggaagac 600  
 ccttcattat tgtggcaggt ttttggcagt gccactggcc tagctcgata ttatccagct 660  
 tcaccatggg ttgataatag tagaactcca aataagattg acctttatga tgtacgcaga 720  
 agaccatggt acatccaagg agctgcatct cctaaagaca tgcttattct ggtggatgtg 780  
 agtggaagtg ttagtggtt gacacttaaa ctgatccgaa catctgtctc cgaaatgtta 840  
 gaaaccctct cagatgatga tttcgtgaat gtagcttcat ttaacagcaa tgctcaggat 900  
 gtaagctgtt ttcagcacct tgtccaagca aatgtaagaa ataaaaaagt gttgaaagac 960  
 gcggtgaata atatcacagc caaaggaatt acagattata agaagggtt tagttttgct 1020  
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 gctaagcaag tccaatggac aaatgtgtac ctggatgcat tggaaactggg acttgtcatt 1380  
 actggaactc ttccggtctt caacataacc ggccaatttg aaaataagac aaacttaaa 1440  
 aaccagctga ttcttggtgt gatgggagta gatgtgtctt tggaagatat taaaagactg 1500  
 acaccacgtt ttacactgtg cccaatggg tattactttg caatcgatcc taatggttat 1560  
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 gatttccttg atgcagagtt agagaatgat attaaagtgg agattcgaaa taagatgatt 1680  
 gatggggaaa gtggagaaaa aacattcaga actctgggta aatctcaaga tgagagatat 1740  
 attgacaaag gaaacaggac atacacatgg acacctgtca atggcacaga ttacagtttg 1800  
 gccttggtat taccaacctc cagttttttac tatataaaag ccaaactaga agagacaata 1860  
 actcaggcca gatcaaaaaa gggcaaaatg aaggattcgg aaaccctgaa gccagataat 1920  
 tttgaagaat ctggctatac attcatagca ccaagagatt actgcaatga cctgaaaata 1980  
 tcggataata aactgaatt tcttttaaat ttcaacgagt ttattgatag aaaaactcca 2040  
 aacaacccat catgtaacgc ggatttgatt aatagagtct tgcttgatgc aggctttaca 2100  
 aatgaacttg tccaaaatta ctggagtaag cagaaaaata tcaaggaggt gaaagcacga 2160  
 tttgttgtga ctgatggtg gattaccaga gtttatccca aagaggctgg agaaaattgg 2220  
 caagaaaacc cagagacata tgaggacagc ttctataaaa ggagcctaga taatgataac 2280  
 tatgttttca ctgctcccta ctttaacaaa agtggacctg gtgcctatga atcgggcatt 2340  
 atggtaagca aagctgtaga aatataatatt caagggaaac ttcttaaaacc tgcagttgtt 2400  
 ggaattaaaa ttgatgtaaa ttcctggata gagaatttca ccaaaacctc aatcagagat 2460  
 ccgtgtgctg gtccagtttg tgactgcaaa agaaacagtg acgtaattga ttgtgtgatt 2520  
 ctggatgatg gtgggtttct tctgatggca aatcatgatg attatactaa tcagattgga 2580  
 agattttttg gagagattga tcccagcttg atgagacacc tgggttaatat atcagtttat 2640  
 gcttttaaca aatcttatga ttatcagtca gtatgtgagc ccggtgctgc accaaaacaa 2700  
 ggagcaggac atcgtcagc atatgtgcc ttagtagcag acatattaca aattggctgg 2760  
 tgggccactg ctgctgctg gtctattcta cagcagtttc tcttgagttt gacctttcca 2820  
 cgactccttg aggcagttga gatggaggat gatgacttca cggcctccct gtccaagcag 2880  
 agctgcatta ctgaacaaac ccagtatttc ttcgataacg acagtaaatc attcagtggt 2940  
 gtattagact gtggaaactg ttccagaatc tttcatggag aaaagcttat gaacaccaac 3000  
 ttaatattca taatggttga gagcaaaggg acatgtccat gtgacacacg actgctcata 3060



caagcggagc agacttctga cgggtccaaat ccttgtagaca tgggtaagca acctagatac 3120  
cgaaaagggc ctgatgtctg ctttgataac aatgtcttgg aggattatac tgactgtggg 3180  
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<210> 13  
<211> 1018  
<212> PRT  
<213> Homo sapiens

<400> 13  
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20 25 30  
Ile Lys Ser Trp Val Asp Lys Met Gln Glu Asp Leu Val Thr Leu Ala  
35 40 45  
Lys Thr Ala Ser Gly Val Asn Gln Leu Val Asp Ile Tyr Glu Lys Tyr  
50 55 60  
Gln Asp Leu Tyr Thr Val Glu Pro Asn Asn Ala Arg Gln Leu Val Glu  
65 70 75 80  
Ile Ala Ala Arg Asp Ile Glu Lys Leu Leu Ser Asn Arg Ser Lys Ala  
85 90 95  
Leu Val Ser Leu Ala Leu Glu Ala Glu Lys Val Gln Ala Ala His Gln  
100 105 110  
Trp Arg Glu Asp Phe Ala Ser Asn Glu Val Val Tyr Tyr Asn Ala Lys  
115 120 125  
Asp Asp Leu Asp Pro Glu Lys Asn Asp Ser Glu Pro Gly Ser Gln Arg  
130 135 140  
Ile Lys Pro Val Phe Ile Glu Asp Ala Asn Phe Gly Arg Gln Ile Ser  
145 150 155 160  
Tyr Gln His Ala Ala Val His Ile Pro Thr Asp Ile Tyr Glu Gly Ser  
165 170 175  
Thr Ile Val Leu Asn Glu Leu Asn Trp Thr Ser Ala Leu Asp Glu Val  
180 185 190  
Phe Lys Lys Asn Arg Glu Glu Asp Pro Ser Leu Leu Trp Gln Val Phe  
195 200 205  
Gly Ser Ala Thr Gly Leu Ala Arg Tyr Tyr Pro Ala Ser Pro Trp Val  
210 215 220  
Asp Asn Ser Arg Thr Pro Asn Lys Ile Asp Leu Tyr Asp Val Arg Arg  
225 230 235 240  
Arg Pro Trp Tyr Ile Gln Gly Ala Ala Ser Pro Lys Asp Met Leu Ile

245										250					255				
Leu	Val	Asp	Val	Ser	Gly	Ser	Val	Ser	Gly	Leu	Thr	Leu	Lys	Leu	Ile				
			260						265					270					
Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe				
			275					280					285						
Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe				
			290					295				300							
Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp				
					310					315					320				
Ala	Val	Asn	Asn	Ile	Thr	Ala	Lys	Gly	Ile	Thr	Asp	Tyr	Lys	Lys	Gly				
				325					330						335				
Phe	Ser	Phe	Ala	Phe	Glu	Gln	Leu	Leu	Asn	Tyr	Asn	Val	Ser	Arg	Ala				
			340						345					350					
Asn	Cys	Asn	Lys	Ile	Ile	Met	Leu	Phe	Thr	Asp	Gly	Gly	Glu	Glu	Arg				
			355					360				365							
Ala	Gln	Glu	Ile	Phe	Asn	Lys	Tyr	Asn	Lys	Asp	Lys	Lys	Val	Arg	Val				
			370				375				380								
Phe	Arg	Phe	Ser	Val	Gly	Gln	His	Asn	Tyr	Glu	Arg	Gly	Pro	Ile	Gln				
					390					395					400				
Trp	Met	Ala	Cys	Glu	Asn	Lys	Gly	Tyr	Tyr	Tyr	Glu	Ile	Pro	Ser	Ile				
				405					410					415					
Gly	Ala	Ile	Arg	Ile	Asn	Thr	Gln	Glu	Tyr	Leu	Asp	Val	Leu	Gly	Arg				
			420					425					430						
Pro	Met	Val	Leu	Ala	Gly	Asp	Lys	Ala	Lys	Gln	Val	Gln	Trp	Thr	Asn				
			435					440				445							
Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu				
			450				455				460								
Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Phe	Glu	Asn	Lys	Thr	Asn	Leu	Lys				
					470					475					480				
Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp				
				485					490					495					
Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr				
			500					505					510						
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln				
			515				520					525							
Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp				
			530				535				540								
Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile				

545		550		555		560
Asp Gly Glu Ser	Gly Glu Lys Thr Phe Arg Thr Leu Val Lys Ser Gln	565	570	575		
Asp Glu Arg Tyr	Ile Asp Lys Gly Asn Arg Thr Tyr Thr Trp Thr Pro	580	585	590		
Val Asn Gly Thr	Asp Tyr Ser Leu Ala Leu Val Leu Pro Thr Tyr Ser	595	600	605		
Phe Tyr Tyr Ile	Lys Ala Lys Leu Glu Glu Thr Ile Thr Gln Ala Arg	610	615	620		
Ser Lys Lys Gly	Lys Met Lys Asp Ser Glu Thr Leu Lys Pro Asp Asn	625	630	635	640	
Phe Glu Glu Ser	Gly Tyr Thr Phe Ile Ala Pro Arg Asp Tyr Cys Asn	645	650	655		
Asp Leu Lys Ile	Ser Asp Asn Asn Thr Glu Phe Leu Leu Asn Phe Asn	660	665	670		
Glu Phe Ile Asp	Arg Lys Thr Pro Asn Asn Pro Ser Cys Asn Ala Asp	675	680	685		
Leu Ile Asn Arg	Val Leu Leu Asp Ala Gly Phe Thr Asn Glu Leu Val	690	695	700		
Gln Asn Tyr Trp	Ser Lys Gln Lys Asn Ile Lys Gly Val Lys Ala Arg	705	710	715	720	
Phe Val Val Thr	Asp Gly Gly Ile Thr Arg Val Tyr Pro Lys Glu Ala	725	730	735		
Gly Glu Asn Trp	Gln Glu Asn Pro Glu Thr Tyr Glu Asp Ser Phe Tyr	740	745	750		
Lys Arg Ser Leu	Asp Asn Asp Asn Tyr Val Phe Thr Ala Pro Tyr Phe	755	760	765		
Asn Lys Ser Gly	Pro Gly Ala Tyr Glu Ser Gly Ile Met Val Ser Lys	770	775	780		
Ala Val Glu Ile	Tyr Ile Gln Gly Lys Leu Leu Lys Pro Ala Val Val	785	790	795	800	
Gly Ile Lys Ile	Asp Val Asn Ser Trp Ile Glu Asn Phe Thr Lys Thr	805	810	815		
Ser Ile Arg Asp	Pro Cys Ala Gly Pro Val Cys Asp Cys Lys Arg Asn	820	825	830		
Ser Asp Val Met	Asp Cys Val Ile Leu Asp Asp Gly Gly Phe Leu Leu	835	840	845		
Met Ala Asn His	Asp Asp Tyr Thr Asn Gln Ile Gly Arg Phe Phe Gly					

850	855	860
Glu Ile Asp Pro Ser Leu Met Arg His Leu Val Asn Ile Ser Val Tyr		
865	870	875 880
Ala Phe Asn Lys Ser Tyr Asp Tyr Gln Ser Val Cys Glu Pro Gly Ala		
	885	890 895
Ala Pro Lys Gln Gly Ala Gly His Arg Ser Ala Tyr Val Pro Ser Val		
	900	905 910
Ala Asp Ile Leu Gln Ile Gly Trp Trp Ala Thr Ala Ala Ala Trp Ser		
	915	920 925
Ile Leu Gln Gln Phe Leu Leu Ser Leu Thr Phe Pro Arg Leu Leu Glu		
	930	935 940
Ala Val Glu Met Glu Asp Asp Asp Phe Thr Ala Ser Leu Ser Lys Gln		
	945	950 955 960
Ser Cys Ile Thr Glu Gln Thr Gln Tyr Phe Phe Asp Asn Asp Ser Lys		
	965	970 975
Ser Phe Ser Gly Val Leu Asp Cys Gly Asn Cys Ser Arg Ile Phe His		
	980	985 990
Gly Glu Lys Leu Met Asn Thr Asn Leu Ile Phe Ile Met Val Glu Ser		
	995	1000 1005
Lys Gly Thr Cys Pro Cys Asp Thr Arg Leu		
1010	1015	

<210> 14  
 <211> 1036  
 <212> PRT  
 <213> Homo sapiens

<400> 14  
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 Ile Lys Ser Trp Val Asp Lys Met Gln Glu Asp Leu Val Thr Leu Ala  
 35 40 45  
 Lys Thr Ala Ser Gly Val Asn Gln Leu Val Asp Ile Tyr Glu Lys Tyr  
 50 55 60  
 Gln Asp Leu Tyr Thr Val Glu Pro Asn Asn Ala Arg Gln Leu Val Glu  
 65 70 75 80  
 Ile Ala Ala Arg Asp Ile Glu Lys Leu Leu Ser Asn Arg Ser Lys Ala  
 85 90 95

Leu	Val	Ser	Leu	Ala	Leu	Glu	Ala	Glu	Lys	Val	Gln	Ala	Ala	His	Gln	
			100					105						110		
Trp	Arg	Glu	Asp	Phe	Ala	Ser	Asn	Glu	Val	Val	Tyr	Tyr	Asn	Ala	Lys	
		115					120					125				
Asp	Asp	Leu	Asp	Pro	Glu	Lys	Asn	Asp	Ser	Glu	Pro	Gly	Ser	Gln	Arg	
	130					135					140					
Ile	Lys	Pro	Val	Phe	Ile	Glu	Asp	Ala	Asn	Phe	Gly	Arg	Gln	Ile	Ser	
145					150					155					160	
Tyr	Gln	His	Ala	Ala	Val	His	Ile	Pro	Thr	Asp	Ile	Tyr	Glu	Gly	Ser	
			165						170					175		
Thr	Ile	Val	Leu	Asn	Glu	Leu	Asn	Trp	Thr	Ser	Ala	Leu	Asp	Glu	Val	
			180					185					190			
Phe	Lys	Lys	Asn	Arg	Glu	Glu	Asp	Pro	Ser	Leu	Leu	Trp	Gln	Val	Phe	
		195					200					205				
Gly	Ser	Ala	Thr	Gly	Leu	Ala	Arg	Tyr	Tyr	Pro	Ala	Ser	Pro	Trp	Val	
	210					215					220					
Asp	Asn	Ser	Arg	Thr	Pro	Asn	Lys	Ile	Asp	Leu	Tyr	Asp	Val	Arg	Arg	
225					230					235					240	
Arg	Pro	Trp	Tyr	Ile	Gln	Gly	Ala	Ala	Ser	Pro	Lys	Asp	Met	Leu	Ile	
			245						250					255		
Leu	Val	Asp	Val	Ser	Gly	Ser	Val	Ser	Gly	Leu	Thr	Leu	Lys	Leu	Ile	
			260					265					270			
Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe	
		275					280					285				
Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe	
	290					295					300					
Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp	
305					310					315					320	
Ala	Val	Asn	Asn	Ile	Thr	Ala	Lys	Gly	Ile	Thr	Asp	Tyr	Lys	Lys	Gly	
				325					330					335		
Phe	Ser	Phe	Ala	Phe	Glu	Gln	Leu	Leu	Asn	Tyr	Asn	Val	Ser	Arg	Ala	
			340					345					350			
Asn	Cys	Asn	Lys	Ile	Ile	Met	Leu	Phe	Thr	Asp	Gly	Gly	Glu	Glu	Arg	
		355					360					365				
Ala	Gln	Glu	Ile	Phe	Asn	Lys	Tyr	Asn	Lys	Asp	Lys	Lys	Val	Arg	Val	
	370					375					380					
Phe	Arg	Phe	Ser	Val	Gly	Gln	His	Asn	Tyr	Glu	Arg	Gly	Pro	Ile	Gln	
385					390					395					400	

Trp Met Ala Cys Glu Asn Lys Gly Tyr Tyr Tyr Glu Ile Pro Ser Ile  
 405 410 415

Gly Ala Ile Arg Ile Asn Thr Gln Glu Tyr Leu Asp Val Leu Gly Arg  
 420 425 430

Pro Met Val Leu Ala Gly Asp Lys Ala Lys Gln Val Gln Trp Thr Asn  
 435 440 445

Val Tyr Leu Asp Ala Leu Glu Leu Gly Leu Val Ile Thr Gly Thr Leu  
 450 455 460

Pro Val Phe Asn Ile Thr Gly Gln Phe Glu Asn Lys Thr Asn Leu Lys  
 465 470 475 480

Asn Gln Leu Ile Leu Gly Val Met Gly Val Asp Val Ser Leu Glu Asp  
 485 490 495

Ile Lys Arg Leu Thr Pro Arg Phe Thr Leu Cys Pro Asn Gly Tyr Tyr  
 500 505 510

Phe Ala Ile Asp Pro Asn Gly Tyr Val Leu Leu His Pro Asn Leu Gln  
 515 520 525

Pro Lys Asn Pro Lys Ser Gln Glu Pro Val Thr Leu Asp Phe Leu Asp  
 530 535 540

Ala Glu Leu Glu Asn Asp Ile Lys Val Glu Ile Arg Asn Lys Met Ile  
 545 550 555 560

Asp Gly Glu Ser Gly Glu Lys Thr Phe Arg Thr Leu Val Lys Ser Gln  
 565 570 575

Asp Glu Arg Tyr Ile Asp Lys Gly Asn Arg Thr Tyr Thr Trp Thr Pro  
 580 585 590

Val Asn Gly Thr Asp Tyr Ser Leu Ala Leu Val Leu Pro Thr Tyr Ser  
 595 600 605

Phe Tyr Tyr Ile Lys Ala Lys Leu Glu Glu Thr Ile Thr Gln Ala Arg  
 610 615 620

Ser Lys Lys Gly Lys Met Lys Asp Ser Glu Thr Leu Lys Pro Asp Asn  
 625 630 635 640

Phe Glu Glu Ser Gly Tyr Thr Phe Ile Ala Pro Arg Asp Tyr Cys Asn  
 645 650 655

Asp Leu Lys Ile Ser Asp Asn Asn Thr Glu Phe Leu Leu Asn Phe Asn  
 660 665 670

Glu Phe Ile Asp Arg Lys Thr Pro Asn Asn Pro Ser Cys Asn Ala Asp  
 675 680 685

Leu Ile Asn Arg Val Leu Leu Asp Ala Gly Phe Thr Asn Glu Leu Val  
 690 695 700

Gln	Asn	Tyr	Trp	Ser	Lys	Gln	Lys	Asn	Ile	Lys	Gly	Val	Lys	Ala	Arg		
705					710					715					720		
Phe	Val	Val	Thr	Asp	Gly	Gly	Ile	Thr	Arg	Val	Tyr	Pro	Lys	Glu	Ala		
				725					730					735			
Gly	Glu	Asn	Trp	Gln	Glu	Asn	Pro	Glu	Thr	Tyr	Glu	Asp	Ser	Phe	Tyr		
			740					745					750				
Lys	Arg	Ser	Leu	Asp	Asn	Asp	Asn	Tyr	Val	Phe	Thr	Ala	Pro	Tyr	Phe		
		755					760					765					
Asn	Lys	Ser	Gly	Pro	Gly	Ala	Tyr	Glu	Ser	Gly	Ile	Met	Val	Ser	Lys		
	770					775					780						
Ala	Val	Glu	Ile	Tyr	Ile	Gln	Gly	Lys	Leu	Leu	Lys	Pro	Ala	Val	Val		
785					790					795					800		
Gly	Ile	Lys	Ile	Asp	Val	Asn	Ser	Trp	Ile	Glu	Asn	Phe	Thr	Lys	Thr		
				805					810					815			
Ser	Ile	Arg	Asp	Pro	Cys	Ala	Gly	Pro	Val	Cys	Asp	Cys	Lys	Arg	Asn		
			820					825					830				
Ser	Asp	Val	Met	Asp	Cys	Val	Ile	Leu	Asp	Asp	Gly	Gly	Phe	Leu	Leu		
		835					840					845					
Met	Ala	Asn	His	Asp	Asp	Tyr	Thr	Asn	Gln	Ile	Gly	Arg	Phe	Phe	Gly		
	850					855					860						
Glu	Ile	Asp	Pro	Ser	Leu	Met	Arg	His	Leu	Val	Asn	Ile	Ser	Val	Tyr		
865					870					875					880		
Ala	Phe	Asn	Lys	Ser	Tyr	Asp	Tyr	Gln	Ser	Val	Cys	Glu	Pro	Gly	Ala		
				885					890					895			
Ala	Pro	Lys	Gln	Gly	Ala	Gly	His	Arg	Ser	Ala	Tyr	Val	Pro	Ser	Val		
			900					905					910				
Ala	Asp	Ile	Leu	Gln	Ile	Gly	Trp	Trp	Ala	Thr	Ala	Ala	Ala	Trp	Ser		
		915					920					925					
Ile	Leu	Gln	Gln	Phe	Leu	Leu	Ser	Leu	Thr	Phe	Pro	Arg	Leu	Leu	Glu		
	930					935					940						
Ala	Val	Glu	Met	Glu	Asp	Asp	Asp	Phe	Thr	Ala	Ser	Leu	Ser	Lys	Gln		
945					950					955					960		
Ser	Cys	Ile	Thr	Glu	Gln	Thr	Gln	Tyr	Phe	Phe	Asp	Asn	Asp	Ser	Lys		
				965					970					975			
Ser	Phe	Ser	Gly	Val	Leu	Asp	Cys	Gly	Asn	Cys	Ser	Arg	Ile	Phe	His		
			980					985					990				
Gly	Glu	Lys	Leu	Met	Asn	Thr	Asn	Leu	Ile	Phe	Ile	Met	Val	Glu	Ser		
		995				1000						1005					

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Thr Ser Asp Gly Pro Asn Pro Cys Asp Met Val Lys  
1025 1030 1035

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<211> 1063

<212> PRT

<213> Homo sapiens

<400> 15

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Ile Lys Ser Trp Val Asp Lys Met Gln Glu Asp Leu Val Thr Leu Ala  
35 40 45

Lys Thr Ala Ser Gly Val Asn Gln Leu Val Asp Ile Tyr Glu Lys Tyr  
50 55 60

Gln Asp Leu Tyr Thr Val Glu Pro Asn Asn Ala Arg Gln Leu Val Glu  
65 70 75 80

Ile Ala Ala Arg Asp Ile Glu Lys Leu Leu Ser Asn Arg Ser Lys Ala  
85 90 95

Leu Val Ser Leu Ala Leu Glu Ala Glu Lys Val Gln Ala Ala His Gln  
100 105 110

Trp Arg Glu Asp Phe Ala Ser Asn Glu Val Val Tyr Tyr Asn Ala Lys  
115 120 125

Asp Asp Leu Asp Pro Glu Lys Asn Asp Ser Glu Pro Gly Ser Gln Arg  
130 135 140

Ile Lys Pro Val Phe Ile Glu Asp Ala Asn Phe Gly Arg Gln Ile Ser  
145 150 155 160

Tyr Gln His Ala Ala Val His Ile Pro Thr Asp Ile Tyr Glu Gly Ser  
165 170 175

Thr Ile Val Leu Asn Glu Leu Asn Trp Thr Ser Ala Leu Asp Glu Val  
180 185 190

Phe Lys Lys Asn Arg Glu Glu Asp Pro Ser Leu Leu Trp Gln Val Phe  
195 200 205

Gly Ser Ala Thr Gly Leu Ala Arg Tyr Tyr Pro Ala Ser Pro Trp Val  
210 215 220



Asp	Asn	Ser	Arg	Thr	Pro	Asn	Lys	Ile	Asp	Leu	Tyr	Asp	Val	Arg	Arg	
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Arg	Pro	Trp	Tyr	Ile	Gln	Gly	Ala	Ala	Ser	Pro	Lys	Asp	Met	Leu	Ile	
				245					250					255		
Leu	Val	Asp	Val	Ser	Gly	Ser	Val	Ser	Gly	Leu	Thr	Leu	Lys	Leu	Ile	
			260					265					270			
Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe	
		275					280					285				
Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe	
	290					295					300					
Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp	
305					310					315					320	
Ala	Val	Asn	Asn	Ile	Thr	Ala	Lys	Gly	Ile	Thr	Asp	Tyr	Lys	Lys	Gly	
				325					330					335		
Phe	Ser	Phe	Ala	Phe	Glu	Gln	Leu	Leu	Asn	Tyr	Asn	Val	Ser	Arg	Ala	
			340					345					350			
Asn	Cys	Asn	Lys	Ile	Ile	Met	Leu	Phe	Thr	Asp	Gly	Gly	Glu	Glu	Arg	
		355					360					365				
Ala	Gln	Glu	Ile	Phe	Asn	Lys	Tyr	Asn	Lys	Asp	Lys	Lys	Val	Arg	Val	
	370					375					380					
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Trp	Met	Ala	Cys	Glu	Asn	Lys	Gly	Tyr	Tyr	Tyr	Glu	Ile	Pro	Ser	Ile	
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Gly	Ala	Ile	Arg	Ile	Asn	Thr	Gln	Glu	Tyr	Leu	Asp	Val	Leu	Gly	Arg	
			420					425					430			
Pro	Met	Val	Leu	Ala	Gly	Asp	Lys	Ala	Lys	Gln	Val	Gln	Trp	Thr	Asn	
		435					440					445				
Val	Tyr	Leu	Asp	Ala	Leu	Glu	Leu	Gly	Leu	Val	Ile	Thr	Gly	Thr	Leu	
	450					455					460					
Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Phe	Glu	Asn	Lys	Thr	Asn	Leu	Lys	
465					470					475					480	
Asn	Gln	Leu	Ile	Leu	Gly	Val	Met	Gly	Val	Asp	Val	Ser	Leu	Glu	Asp	
				485				490						495		
Ile	Lys	Arg	Leu	Thr	Pro	Arg	Phe	Thr	Leu	Cys	Pro	Asn	Gly	Tyr	Tyr	
			500					505					510			
Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln	
		515					520					525				

Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp		
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Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile		
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Asp	Glu	Arg	Tyr	Ile	Asp	Lys	Gly	Asn	Arg	Thr	Tyr	Thr	Trp	Thr	Pro		
			580					585					590				
Val	Asn	Gly	Thr	Asp	Tyr	Ser	Leu	Ala	Leu	Val	Leu	Pro	Thr	Tyr	Ser		
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Phe	Glu	Glu	Ser	Gly	Tyr	Thr	Phe	Ile	Ala	Pro	Arg	Asp	Tyr	Cys	Asn		
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			660					665					670				
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		675					680					685					
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Lys	Arg	Ser	Leu	Asp	Asn	Asp	Asn	Tyr	Val	Phe	Thr	Ala	Pro	Tyr	Phe		
		755					760					765					
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770						775					780						
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785					790					795					800		
Gly	Ile	Lys	Ile	Asp	Val	Asn	Ser	Trp	Ile	Glu	Asn	Phe	Thr	Lys	Thr		
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Ser	Ile	Arg	Asp	Pro	Cys	Ala	Gly	Pro	Val	Cys	Asp	Cys	Lys	Arg	Asn		
			820					825					830				

Ser Asp Val Met Asp Cys Val Ile Leu Asp Asp Gly Gly Phe Leu Leu  
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 Met Ala Asn His Asp Asp Tyr Thr Asn Gln Ile Gly Arg Phe Phe Gly  
 850 855 860  
 Glu Ile Asp Pro Ser Leu Met Arg His Leu Val Asn Ile Ser Val Tyr  
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 Ala Phe Asn Lys Ser Tyr Asp Tyr Gln Ser Val Cys Glu Pro Gly Ala  
 885 890 895  
 Ala Pro Lys Gln Gly Ala Gly His Arg Ser Ala Tyr Val Pro Ser Val  
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 Ala Asp Ile Leu Gln Ile Gly Trp Trp Ala Thr Ala Ala Ala Trp Ser  
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 Ile Leu Gln Gln Phe Leu Leu Ser Leu Thr Phe Pro Arg Leu Leu Glu  
 930 935 940  
 Ala Val Glu Met Glu Asp Asp Asp Phe Thr Ala Ser Leu Ser Lys Gln  
 945 950 955 960  
 Ser Cys Ile Thr Glu Gln Thr Gln Tyr Phe Phe Asp Asn Asp Ser Lys  
 965 970 975  
 Ser Phe Ser Gly Val Leu Asp Cys Gly Asn Cys Ser Arg Ile Phe His  
 980 985 990  
 Gly Glu Lys Leu Met Asn Thr Asn Leu Ile Phe Ile Met Val Glu Ser  
 995 1000 1005  
 Lys Gly Thr Cys Pro Cys Asp Thr Arg Leu Leu Ile Gln Ala Glu Gln  
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<210> 16  
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 <212> PRT  
 <213> Homo sapiens

<400> 16  
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Leu Leu Ile Gly Pro Ser Ser Glu Glu Pro Phe Pro Ser Ala Val Thr

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Gln	Asp	Leu	Tyr	Thr	Val	Glu	Pro	Asn	Asn	Ala	Arg	Gln	Leu	Val	Glu
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Ile	Ala	Ala	Arg	Asp	Ile	Glu	Lys	Leu	Leu	Ser	Asn	Arg	Ser	Lys	Ala
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Leu	Val	Ser	Leu	Ala	Leu	Glu	Ala	Glu	Lys	Val	Gln	Ala	Ala	His	Gln
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Trp	Arg	Glu	Asp	Phe	Ala	Ser	Asn	Glu	Val	Val	Tyr	Tyr	Asn	Ala	Lys
		115					120					125			
Asp	Asp	Leu	Asp	Pro	Glu	Lys	Asn	Asp	Ser	Glu	Pro	Gly	Ser	Gln	Arg
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Ile	Lys	Pro	Val	Phe	Ile	Glu	Asp	Ala	Asn	Phe	Gly	Arg	Gln	Ile	Ser
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Tyr	Gln	His	Ala	Ala	Val	His	Ile	Pro	Thr	Asp	Ile	Tyr	Glu	Gly	Ser
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Thr	Ile	Val	Leu	Asn	Glu	Leu	Asn	Trp	Thr	Ser	Ala	Leu	Asp	Glu	Val
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Phe	Lys	Lys	Asn	Arg	Glu	Glu	Asp	Pro	Ser	Leu	Leu	Trp	Gln	Val	Phe
		195					200					205			
Gly	Ser	Ala	Thr	Gly	Leu	Ala	Arg	Tyr	Tyr	Pro	Ala	Ser	Pro	Trp	Val
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Asp	Asn	Ser	Arg	Thr	Pro	Asn	Lys	Ile	Asp	Leu	Tyr	Asp	Val	Arg	Arg
	225					230					235				240
Arg	Pro	Trp	Tyr	Ile	Gln	Gly	Ala	Ala	Ser	Pro	Lys	Asp	Met	Leu	Ile
				245					250					255	
Leu	Val	Asp	Val	Ser	Gly	Ser	Val	Ser	Gly	Leu	Thr	Leu	Lys	Leu	Ile
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Arg	Thr	Ser	Val	Ser	Glu	Met	Leu	Glu	Thr	Leu	Ser	Asp	Asp	Asp	Phe
		275					280					285			
Val	Asn	Val	Ala	Ser	Phe	Asn	Ser	Asn	Ala	Gln	Asp	Val	Ser	Cys	Phe
	290					295					300				
Gln	His	Leu	Val	Gln	Ala	Asn	Val	Arg	Asn	Lys	Lys	Val	Leu	Lys	Asp
	305					310					315				320
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Pro	Val	Phe	Asn	Ile	Thr	Gly	Gln	Phe	Glu	Asn	Lys	Thr	Asn	Leu	Lys																																
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Phe	Ala	Ile	Asp	Pro	Asn	Gly	Tyr	Val	Leu	Leu	His	Pro	Asn	Leu	Gln																																
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Pro	Lys	Asn	Pro	Lys	Ser	Gln	Glu	Pro	Val	Thr	Leu	Asp	Phe	Leu	Asp																																
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Ala	Glu	Leu	Glu	Asn	Asp	Ile	Lys	Val	Glu	Ile	Arg	Asn	Lys	Met	Ile																																
			545														550													555			560														
Asp	Gly	Glu	Ser	Gly	Glu	Lys	Thr	Phe	Arg	Thr	Leu	Val	Lys	Ser	Gln																																
			565														570													575																	
Asp	Glu	Arg	Tyr	Ile	Asp	Lys	Gly	Asn	Arg	Thr	Tyr	Thr	Trp	Thr	Pro																																
			580														585													590																	
Val	Asn	Gly	Thr	Asp	Tyr	Ser	Leu	Ala	Leu	Val	Leu	Pro	Thr	Tyr	Ser																																
			595														600													605																	
Phe	Tyr	Tyr	Ile	Lys	Ala	Lys	Leu	Glu	Glu	Thr	Ile	Thr	Gln	Ala	Arg																																
			610														615													620																	
Ser	Lys	Lys	Gly	Lys	Met	Lys	Asp	Ser	Glu	Thr	Leu	Lys	Pro	Asp	Asn																																

625		630		635		640
Phe Glu Glu Ser Gly Tyr Thr Ph Ile Ala Pro Arg Asp Tyr Cys Asn	645		650		655	
Asp Leu Lys Ile Ser Asp Asn Asn Thr Glu Phe Leu Leu Asn Phe Asn	660		665		670	
Glu Phe Ile Asp Arg Lys Thr Pro Asn Asn Pro Ser Cys Asn Ala Asp	675		680		685	
Leu Ile Asn Arg Val Leu Leu Asp Ala Gly Phe Thr Asn Glu Leu Val	690		695		700	
Gln Asn Tyr Trp Ser Lys Gln Lys Asn Ile Lys Gly Val Lys Ala Arg	705		710		715	720
Phe Val Val Thr Asp Gly Gly Ile Thr Arg Val Tyr Pro Lys Glu Ala	725		730		735	
Gly Glu Asn Trp Gln Glu Asn Pro Glu Thr Tyr Glu Asp Ser Phe Tyr	740		745		750	
Lys Arg Ser Leu Asp Asn Asp Asn Tyr Val Phe Thr Ala Pro Tyr Phe	755		760		765	
Asn Lys Ser Gly Pro Gly Ala Tyr Glu Ser Gly Ile Met Val Ser Lys	770		775		780	
Ala Val Glu Ile Tyr Ile Gln Gly Lys Leu Leu Lys Pro Ala Val Val	785		790		795	800
Gly Ile Lys Ile Asp Val Asn Ser Trp Ile Glu Asn Phe Thr Lys Thr	805		810		815	
Ser Ile Arg Asp Pro Cys Ala Gly Pro Val Cys Asp Cys Lys Arg Asn	820		825		830	
Ser Asp Val Met Asp Cys Val Ile Leu Asp Asp Gly Gly Phe Leu Leu	835		840		845	
Met Ala Asn His Asp Asp Tyr Thr Asn Gln Ile Gly Arg Phe Phe Gly	850		855		860	
Glu Ile Asp Pro Ser Leu Met Arg His Leu Val Asn Ile Ser Val Tyr	865		870		875	880
Ala Phe Asn Lys Ser Tyr Asp Tyr Gln Ser Val Cys Glu Pro Gly Ala	885		890		895	
Ala Pro Lys Gln Gly Ala Gly His Arg Ser Ala Tyr Val Pro Ser Val	900		905		910	
Ala Asp Ile Leu Gln Ile Gly Trp Trp Ala Thr Ala Ala Ala Trp Ser	915		920		925	
Ile Leu Gln Gln Phe Leu Leu Ser Leu Thr Phe Pro Arg Leu Leu Glu						

930	935	940
Ala Val Glu Met Glu Asp Asp Asp Phe Thr Ala Ser Leu Ser Lys Gln 945 950 955 960		
Ser Cys Ile Thr Glu Gln Thr Gln Tyr Phe Phe Asp Asn Asp Ser Lys 965 970 975		
Ser Phe Ser Gly Val Leu Asp Cys Gly Asn Cys Ser Arg Ile Phe His 980 985 990		
Gly Glu Lys Leu Met Asn Thr Asn Leu Ile Phe Ile Met Val Glu Ser 995 1000 1005		
Lys Gly Thr Cys Pro Cys Asp Thr Arg Leu Leu Ile Gln Ala Glu Gln 1010 1015 1020		
Thr Ser Asp Gly Pro Asn Pro Cys Asp Met Val Lys Gln Pro Arg Tyr 1025 1030 1035 1040		
Arg Lys Gly Pro Asp Val Cys Phe Asp Asn Asn Val Leu Glu Asp Tyr 1045 1050 1055		
Thr Asp Cys Gly Gly Val Ser Gly Leu Asn Pro Ser Leu Trp Tyr Ile 1060 1065 1070		
Ile Gly Ile Gln Phe Leu Leu Leu Trp Leu Val Ser Gly Ser Thr His 1075 1080 1085		
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 <212> DNA  
 <213> Homo sapiens

<400> 17

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